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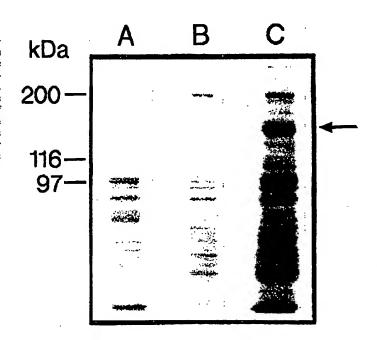
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(54) Title: AN INTEGRIN HETERODIMER AND A SUBUNIT THEREOF

A recombinant or isolated integrin heterodimer comprising a novel subunit a 10 in association with a subunit β is described. The α 10 integrin may be purified from bovine chondrocytes on a collagen-type-II affinity column. The integrin or the subunit $\alpha 10$ can be used as marker or target of all types of cells, e.g. of chondrocytes, osteoblasts and fibroblasts. The integrin or subunit a 10 thereof can be used as marker or target in different physiological or therapeutic methods. They can also be used as active ingredients in pharmaceutical compositions and vaccines.



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AN INTEGRIN HETERODIMER AND A SUBUNIT THEREOF

FIELD OF THE INVENTION

The present invention relates to a recombinant or isolated integrin heterodimer comprising a subunit $\alpha 10$ and a subunit β , the subunit $\alpha 10$ thereof, homologues and fragments of said integrin and of said subunit $\alpha 10$ having similar biological activity, processes of producing the same, polynucleotides and oligonucleotides encoding the same, vectors and cells comprising the same, binding entities binding specifically to the same, and the use of the same.

BACKGROUND OF THE INVENTION

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The integrins are a large family of transmembrane glycoproteins that mediate cell-cell and cell-matrix 15 interactions (1-5). All known members of this superfamily are non-covalently associated heterodimers composed of an α - and a β -subunit. At present, 8 β -subunits (β 1- β 8) (6) and 16 α -subunits (α 1- α 9, α v, α M, α L, α X, α IIb, α E and 20 αD) have been characterized (6-21), and these subunits associate to generate more than 20 different integrins. The β 1-subunit has been shown to associate with ten different α -subunits, $\alpha 1-\alpha 9$ and αv , and to mediate interactions with extracellular matrix proteins such as colla-25 gens, laminins and fibronectin. The major collagen binding integrins are $\alpha 1\beta 1$ and $\alpha 2\beta 1$ (22-25). The integrins $\alpha 3\beta 1$ and $\alpha 9\beta 1$ have also been reported to interact with collagen (26,27) although this interaction is not well understood (28). The extracellular N-terminal regions of the α and β integrin subunits are important in the bind-30 ing of ligands (29,30). The N-terminal region of the α-subunits is composed of a seven-fold repeated sequence (12,31) containing FG and GAP consensus sequences. The repeats are predicted to fold into a β -propeller domain

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(32) with the last three or four repeats containing putative divalent cation binding sites. The α -integrin subunits $\alpha 1$, $\alpha 2$, αD , αE , αL , αM and αX contain a ~200 amino acid inserted domain, the I-domain (A-domain), which shows similarity to sequences in von Willebrand factor, cartilage matrix protein and complement factors C2 and B (33,34). The I-domain is localized between the second and third FG-GAP repeats, it contains a metal ion-dependent adhesion site (MIDAS) and it is involved in binding of ligands (35-38).

Chondrocytes, the only type of cells in cartilage, express a number of different integrins including $\alpha 1\beta 1$, $\alpha 2\beta 1$, $\alpha 3\beta 1$, $\alpha 5\beta 1$, $\alpha 6\beta 1$, $\alpha v\beta 3$, and $\alpha v\beta 5$ (39-41). It has been shown that $\alpha 1\beta 1$ and $\alpha 2\beta 1$ mediate chondrocyte interactions with collagen type II (25) which is one of the major components in cartilage. It has also been shown that $\alpha 2\beta 1$ is a receptor for the cartilage matrix protein chondroadherin (42).

20 SUMMARY OF THE INVENTION

The present invention relates to a novel collagen type II binding integrin, comprising a subunit $\alpha 10$ in association with a subunit β , especially subunit $\beta 1$, but also other β -subunits may be contemplated. In preferred embodiments, this integrin has been isolated from human or bovine articular chondrocytes, and human chondrosarcoma cells.

The invention also encompasses integrin homologues of said integrin, isolated from other species, such as bovine integrin heterodimer comprising a subunit $\alpha 10$ in association with a subunit β , preferably $\beta 1$, as well as homologues isolated from other types of human cells or from cells originating from other species.

The present invention relates in particular to a recombinant or isolated integrin subunit $\alpha 10$ comprising the amino acid sequence shown in SEQ ID No. 1 or SEQ ID No. 2, and homologues and or fragments thereof having the

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same biological activity.

The invention further relates to a process of producing a recombinant integrin subunit $\alpha 10$ comprising the amino acid sequence shown in SEQ ID No. 1 or SEQ ID No. 2, or homologues or fragments thereof having similar biological activity, which process comprises the steps of

- a) isolating a polynucleotide comprising a nucleotide sequence coding for a integrin subunit $\alpha 10$, or homologues or fragments thereof having similar biological activity,
- b) constructing an expression vector comprising the isolated polynucleotide,
- c) transforming a host cell with said expression vector,
- d) culturing said transformed host cell in a culture medium under conditions suitable for expression of integrin subunit $\alpha 10$, or homologues or fragments thereof having similar biological activity, in said transformed host cell, and, optionally,
 - e) isolating the integrin subunit $\alpha 10$, or homologues or fragments thereof having the same biological activity, from said transformed host cell or said culture medium.

The integrin subunit $\alpha 10$, or homologues or fragments thereof having the same biological activity, can also be provided by isolation from a cell in which they are naturally present.

The invention also relates to an isolated polynucleotide comprising a nucleotide coding for a integrin subunit $\alpha 10$, or homologues or fragments thereof having similar biological activity, which polynucleotide comprises the nucleotide sequence shown in SEQ ID No. 1 or SEQ ID No. 2, or parts thereof.

The invention further relates to an isolated polynucleotide or oligonucleotide which hybridises to a DNA or RNA encoding an integrin subunit $\alpha 10$, having the amino acid sequence shown in SEQ ID No. 1 or SEQ ID No. 2, or homologues or fragments thereof, wherein said polyoligo-

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nucleotide or oligonucleotide fails to hybridise to a DNA or RNA encoding the integrin subunit α 1.

The invention relates in a further aspect to vectors comprising the above polynucleotides, and to cells containing said vectors and cells that have polynucleotides or oligonycleotides as shown in SEQ ID No. 1 or 2 integrated in their genome.

The invention also relates to binding entities having the capability of binding specifically to the integrin subunit $\alpha 10$ or to homologues or fragments thereof, such as proteins, peptides, carbohydrates, lipids, natural ligands, polyclonal antibodies or monoclonal antibodies.

In a further aspect, the invention relates to a recombinant or isolated integrin heterodimer comprising a subunit $\alpha 10$ and a subunit β , in which the subunit $\alpha 10$ comprises the amino acid sequence shown in SEQ ID No. 1 or SEQ ID No. 2, or homologues or fragments thereof having similar biological activity.

In a preferred embodiment thereof, the subunit β is β 1.

The invention also relates to a process of producing a recombinant integrin heterodimer comprising a subunit $\alpha 10$ and a subunit β , in which the subunit $\alpha 10$ comprises the amino acid sequence shown in SEQ ID No. 1 or SEQ ID No. 2, which process comprises the steps of

- a) isolating one polynucleotide comprising a nucleotide sequence coding for a subunit $\alpha 10$ of an integrin heterodimer and, optionally, another polynucleotide comprising a nucleotide sequence coding for a subunit β of an integrin heterodimer, or for homologues or fragments thereof having similar biological activity,
- b) constructing an expression vector comprising said isolated polynucleotide coding for said subunit $\alpha 10$ in combination with an expression vector comprising said isolated nucleotide coding for said subunit β ,

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- c) transforming a host cell with said expression vectors.
- d) culturing said transformed host cell in a culture medium under conditions suitable for expression of an integrin heterodimer comprising a subunit all and a subunit β , or homologues or fragments thereof similar biological activity, in said transformed host cell, and, optionally,
- e) isolating the integrin heterodimer comprising a subunit $\alpha 10$ and a subunit β , or homologues or fragments thereof having the same biological activity, from said transformed host cell or said culture medium.

The integrin heterodimer, or homologues or fragments thereof having similar biological activity, can also be provided by isolation from a cell in which they are naturally present.

The invention further relates to a cell containing a first vector, said first vector comprising a polynucleotide coding for a subunit alo of an integrin heterodimer, 20 or for homologues or parts thereof having similar biological activity, which polynucleotide comprises the nucleotide sequence shown in SEQ ID No. 1 or SEQ ID No. 2 or parts thereof, and, optionally, a second vector, said second vector comprising a polynucleotide coding for a subunit β of an integrin heterodimer, or for homologues or fragments thereof.

In still another aspect, the invention relates to binding entities having the capability of binding specifically to the integrin heterodimer comprising a subunit α 10 and a subunit β , or to homologues or fragments thereof having similar biological activity, preferably wherein the subunit β is β 1. Preferred binding entities are proteins, peptides, carbohydrates, lipids, natural ligands, polyclonal antibodies and monoclonal antibodies.

In a further aspect, the invention relates to a fragment of the integrin subunit $\alpha 10$, which fragment is a peptide chosen from the group comprising peptides of

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the cytoplasmic domain, the I-domain and the spliced domain.

In one embodiment, said fragment is a peptide comprising the amino acid sequence KLGFFAHKKIPEEEKREEKLEQ.

In another embodiment, said fragment comprises the amino acid sequence from about amino acid no. 952 to about amino acid no. 986 of SEQ ID No. 1.

In a further embodiment, said fragment comprises the amino acid sequence from about amino acid No. 140 to about amino acid No. 337 in SEO ID No. 1.

Another embodiment of the invention relates to a polynucleotide or oligonucleotide coding for a fragment of the human integrin subunit $\alpha 10$. In one embodiment this polynucleotide of oligonucleotide codes for a fragment which is a peptide chosen from the group comprising peptides of the cytoplasmic domain, the I-domain and the spliced domain. In further embodiments the polynucleotide or oligonucleotide codes for the fragments defined above.

The invention also relates to binding entities having the capability of binding specifically to a fragment of the integrin subunit $\alpha 10$ as defined above.

The invention also relates to a process of using an integrin subunit $\alpha 10$ comprising the amino acid sequence shown in SEQ ID No. 1 or SEQ ID No. 2, or an integrin heterodimer comprising said subunit $\alpha 10$ and a subunit β , or a homologue or fragment of said integrin or subunit having similar biological activity, as a marker or target molecule of cells or tissues expressing said integrin subunit $\alpha 10$, which cells or tissues are of animal including human origin.

In an embodiment of this process the fragment is a peptide chosen from the group comprising peptides of the cytoplasmic domain, the I-domain and the spliced domain.

In further embodiments of said process the fragment is a peptide comprising the amino acid sequence KLGFFAHKKIPEEEKREEKLEQ, or a fragment comprising the amino acid sequence from about amino acid No. 952 to

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about amino acid No. 986 of SEQ ID No. 1, or a fragment comprising the amino acid sequence from about amino acid no. 140 to about amino acid no. 337 of SEQ ID no. 1.

The subunit β is preferably $\beta 1$. The cells are preferably chosen from the group comprising chondrocytes, smooth muscle cells, endothelial cells, osteoblasts and fibroblasts.

Said process may be used during pathological conditions involving said subunit $\alpha 10$, such as pathological conditions comprising damage of cartilage, or comprising trauma, rheumatoid arthritis and osteoarthritis.

Said process may be used for detecting the formation of cartilage during embryonal development, or for detecting physiological or therapeutic reparation of cartilage.

Said process may also be used for selection and analysis, or for sorting, isolating or purification of chondrocytes.

A further embodiment of said process is a process for detecting regeneration of cartilage or chondrocytes during transplantation of cartilage or chondrocytes.

A still further embodiment of said process is a process for in vitro studies of differentiation of chondrocytes.

The invention also comprises a process of using binding entities having the capability of binding specifically to an integrin subunit $\alpha 10$ comprising the amino acid sequence shown in SEQ ID No. 1 or SEQ ID No. 2, or an integrin heterodimer comprising said subunit $\alpha 10$ and a subunit β , or to homologues or fragments thereof having similar biological activity, as markers or target molecules of cells or tissues expressing said integrin subunit $\alpha 10$, which cells or tissues are of animal including human origin.

The fragment in said process may be a peptide chosen from the group comprising peptides of the cytoplasmic domain, the I-domain and the spliced domain. In preferred embodiments said fragment is a peptide comprising the

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amino acid sequence KLGFFAHKKIPEEEKREEKLEQ, or a fragment comprising the amino acid sequence from about amino acid No. 952 to about amino acid No. 986 of SEQ ID No. 1, or a fragment comprising the amino acid sequence from about amino acid No. 140 to about amino acid no. 337 of SEQ ID No. 1.

The process may also be used for detecting the presence of an integrin subunit $\alpha 10$ comprising the amino acid sequence shown in SEQ ID No. 1 or SEQ ID No. 2, or of an integrin heterodimer comprising said subunit $\alpha 10$ and a subunit β , or of homologues or fragments thereof having similar biological activity.

In a further embodiment said process is a process for determining the differentiation-state of cells during embryonic development, angiogenesis, or development of cancer.

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In a still further embodiment this process is a process for detecting the presence of an integrin subunit α 10, or of a homologue or fragment of said integrin subunit having similar biological activity, on cells, 20 whereby a polynucleotide or oligonucleotide chosen from the group comprising a polynucleotide or oligonucleotide chosen from the nucelotide sequence shown in SEQ ID No. 1 is used as a marker under hybridisation conditions where-25 in said polynucleotide or oligonucleotide fails to hybridise to a DNA or RNA encoding an integrin subunit α1. Said cells may be chosen from the group comprising chondrocytes, smooth muscle cells, endothelial cells, osteoblasts and fibroblasts. Said integrin fragment may be a peptide chosen from the group comprising peptides of the cytoplasmic domain, the I-domain and the spliced domain, such as a peptide comprising the amino acid sequence KLGFFAHKKIPEEEKREEKLEQ, or a fragment comprising the amino acid sequence from about amino acid no. 952 to about amino acid no. 986 of SEQ ID No. 1, or a fragment 35 comprising the amino acid sequence from about amino acid No. 140 to about amino acid no. 337 of SEO ID No. 1.

In a still further embodiment the process is a process for determining the differentiation-state of cells during development, in pathological conditions, in tissue regeneration or in therapeutic and physiological reparation of cartilage. The pathological conditions may be any pathological conditions involving the integrin subunit $\alpha 10$, such as rheumatoid arthritis, osteoarthrosis or cancer. The cells may be chosen from the group comprising chondrocytes, smooth muscle cells, endothelial cells, osteoblasts and fibroblasts.

The invention also relates to a process for determining the differentiation-state of cells during development, in pathological conditions, in tissue regeneration and in therapeutic and physiological reparation of car-15 tilage, whereby a polynucleotide or oligonucleotide chosen from the nucleotide sequence shown in SEQ ID No. 1 is used as a marker under hybridisation conditions wherein said polynucleotide or oligonucleotide fails to hybridise to a DNA or RNA encoding an integrin subunit α1. Embodiments of this aspect comprise a process, where-20 by said polynucleotide or oligonucleotide is a polynucleotide or oligonucleotide coding for a peptide chosen from the group comprising peptides of the cytoplasmic domain, the I-domain and the spliced domain, 25 such as a polynucleotide or oligonucleotide coding for a peptide comprising the amino acid sequence KLGFFAHKKIPEEEKREEKLEQ, or comprising the amino acid sequence from about amino acid No. 952 to about amino acid no. 986 of SEQ ID No. 1, or the amino acid sequence from about amino acid No. 140 to about amino acid No. 337 30 of SEQ ID No. 1. Said pathological conditions may be any pathological conditions involving the integrin subunit α10, such as rheumatoid arthritis, osteoarthrosis or cancer, or atherosclerosis or inflammation. Said cells 35 may be chosen from the group comprising chondrocytes, smooth muscle cells, endothelial cells, osteoblasts and fibroblasts.

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In a further aspect the invention relates to a pharmaceutical composition comprising as an active ingredient a pharmaceutical agent or an antibody which is capable of using an integrin heterodimer comprising a subunit $\alpha 10$ and a subunit β , or the subunit $\alpha 10$ thereof, or a homologue or fragment of said integrin or subunit $\alpha 10$ having similar biological activity, as a target molecule. An embodiment of said pharmaceutical composition is intended for use in stimulating, inhibiting or blocking the formation of cartilage, bone or blood vessels. A further embodiment comprises a pharmaceutical composition for use in preventing adhesion between tendon/ligaments and the surrounding tissue after infection, inflammation and after surgical intervention where adhesion impairs the function of the tissue.

The invention is also related to a vaccine comprising as an active ingredient an integrin heterodimer comprising a subunit $\alpha 10$ and a subunit β , or the subunit $\alpha 10$ thereof, or a homologue or fragment of said integrin or subunit $\alpha 10$, or DNA or RNA coding for said integrin subunit $\alpha 10$.

A further aspect of the invention is related to the use of the integrin subunit $\alpha 10$ as defined above as a marker or target in transplantation of cartilage or chondrocytes.

A still further aspect of the invention is related to a method of using binding entities having the capability of binding specifically to an integrin subunit $\alpha 10$ comprising the amino acid sequence shown in SEQ ID No. 1 or SEQ ID No. 2, or an integrin heterodimer comprising said subunit $\alpha 10$ and a subunit β , or to homologues or fragments thereof having similar biological activity, for promoting adhesion of chondrocytes and/or osteoblasts to surfaces of implants to stimulate osseointegration.

35 The invention is also related to the use of an integrin subunit $\alpha 10$ or an integrin heterodimer comprising said subunit $\alpha 10$ and a subunit β as a target for anti-

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adhesive drugs or molecules in tendon, ligament, skeletal muscle or other tissues where adhesion impairs the function of the tissue.

The invention also relates to a method of stimulating, inhibiting or blocking the formation of cartilage or bone, comprising administration to a subject a suitable amount of a pharmaceutical agent or an antibody which is capable of using an integrin heterodimer comprising a subunit $\alpha 10$ and a subunit β , or the subunit $\alpha 10$ thereof, or a homologue or fragment of said integrin or subunit $\alpha 10$ having similar biological activity, as a target molecule.

In another embodiment the invention is related to a method of preventing adhesion between tendon/ligaments and the surrounding tissue after infection, inflammation and after surgical intervention where adhesion impairs the function of the tissue, comprising administration to a subject a suitable amount of a pharmaceutical agent or an antibody which is capable of using a integrin heterodimer comprising a subunit $\alpha 10$ and a subunit β , or the subunit $\alpha 10$ thereof, or a homologue or fragment of said integrin or subunit $\alpha 10$ having similar biological activity, as a target molecule.

The invention also relates to a method of stimulating extracellular matrix synthesis and repair by activation or blockage of an integrin heterodimer comprising a subunit $\alpha 10$ and a subunit β , or of the subunit $\alpha 10$ thereof, or of a homologue or fragment of said integrin or subunit $\alpha 10$ having similar biological activity.

In a further aspect the invention relates to a method of in vitro detecting the presence of integrin binding entities, comprising interaction of an integrin heterodimer comprising a subunit $\alpha 10$ and a subunit β , or the subunit $\alpha 10$ thereof, or a homologue or fragment of said integrin or subunit, with a sample, thereby causing said integrin, subunit $\alpha 10$, or homologue or fragment thereof having similar biological activity, to modulate

the binding to its natural ligand or other integrin binding proteins present in said sample.

The invention also relates to a method of in vitro studying consequences of the interaction of a human heterodimer integrin comprising a subunit $\alpha 10$ and a subunit β , or the subunit $\alpha 10$ thereof, or a homologue or fragment of said integrin or subunit, with an integrin binding entity and thereby initiate a cellular reaction. Said consequences may be measured as alterations in cellular functions.

A still further aspect of the inventions relates to a method of using DNA or RNA encoding an integrin subunit $\alpha 10$ or homologues or fragments thereof as a molecular target. In an embodiment of this aspect, a polynucleotide or oligonucleotide hybridises to the DNA or RNA encoding an integrin subunit $\alpha 10$ or homologues or fragments thereof, whereby said polynucleotide or oligonucleotide fails to hybridise to a DNA or RNA encoding en integrin subunit $\alpha 1$.

The invention also relates to a method of using a human heterodimer integrin comprising a subunit $\alpha 10$ and a subunit β , or the subunit $\alpha 10$ thereof, or a homologue or fragment of said integrin or subunit, or a DNA or RNA encoding an integrin subunit $\alpha 10$ or homologues or fragments thereof, as a marker or target molecule during angiogenesis.

BRIEF DESCRIPTION OF THE FIGURES

Fig.1 Affinity purification of the $\alpha 10$ integrin subunit on collagen type II-Sepharose.

30 Fig. 2. Amino acid sequences of peptides from the bovine $\alpha 10$ integrin subunit.

Fig. 3a. Affinitypurification and immunoprecipitation of the integrin subunit $\alpha 10$ from bovine chondrocytes.

Fig. 3b. Affinitypurification and immunoprecipitation of the integrin subunit $\alpha 10$ from human chondrocytes.

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Fig. 3c. Affinitypurification and immunoprecipitation of the integrin subunit $\alpha 10$ from human chondrosarcoma cells.

- Fig. 4. A 900 bp PCR-fragment corresponding to the bovine integrin subunit $\alpha 10$
 - Fig. 5. Schematic map of the three overlapping $\alpha 10\,$ clones.
 - Fig. 6. Nucleotide sequence and deduced amino acid sequence of the human $\alpha 10$ integrin subunit.
- 10 Fig. 7. Northern blot of integrin α10 mRNA.
 - Fig. 8 Immunoprecipitation of the $\alpha 10$ integrin subunit from human chondrocytes using antibodies against the cytoplasmic domain of $\alpha 10$ (a). Western blot of the $\alpha 10$ associated β -chain (b).
- 15 Fig. 9. Immunostaining of $\alpha 10$ integrin in human articular cartilage.
 - Fig. 10 Immunostaining of $\alpha 10$ integrin in 3 day mouse limb cartilage.
- Fig 11. Immunostaining of α 10 integrin in 13.5 day 20 mouse embryo.
 - Fig 12. Hybridisation of $\alpha 10~\text{mRNA}$ in various human tissues.
- Fig. 13 Immunostaining of fascia around tendon (a), skeletal muscle (b) and heart valves (c) in 3 day mouse 25 limb.
 - Fig. 14. PCR fragments corresponding to $\alpha 10$ integrin subunit from human chondrocytes, human endothelial cells, human fibroblasts and rat tendon.
- Fig 15. Partial genomic nucleotide sequence of the 30 human integrin subunit $\alpha 10\,.$
 - Fig 16. Upregulation of $\alpha 10$ integrin subunit in chondrocytes cultured in alginate.
 - Fig 17. Immunoprecipitation of the $\alpha 10$ integrin subunit from human smooth muscle cells

DETAILED DESCRIPTION OF THE INVENTION

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The present invention demonstrate that human and

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bovine chondrocytes express a novel, collagen type II-binding integrin in the β 1-family. An earlier study presented some evidence for that human chondrosarcoma cells also express this integrin (25). Immunoprecipitation experiments using antibodies against the integrin subunit β 1 revealed that this novel α -integrin subunit had an apparent molecular weight (M_r) of approximately 160 kDa under reducing conditions, and was slightly larger than the $\alpha 2$ integrin subunit. To isolate this α-subunit collagen type II-binding proteins were affinity 10 purified from bovine chondrocytes. The chondrocyte lysate was first applied to a fibronectin-Sepharose precolumn and the flow through was then applied to a collagen type II-Sepharose column. A protein with Mr of approximately 160 kD was specifically eluted with EDTA from the colla-15 gen column but not from the fibronectin column. The $M_{\rm r}$ of this protein corresponded with the M_r of the unidentified β1-related integrin subunit. The 160 kD protein band was excised from the SDS-PAGE gel, digested with trypsin and the amino acid sequences of the isolated peptides were 20 analysed.

Primers corresponding to isolated peptides amplified a 900 bp PCR-fragment from bovine cDNA which was cloned, sequenced and used for screening of a human articular chondrocyte $\lambda ZapII$ cDNA library to obtain the human integrin α -subunit homologue. Two overlapping clones, hcl and hc2 were isolated, subcloned and sequenced. These clones contained 2/3 of the nucleotide sequence including the 3' end of the cDNA. A third clone which contained the 5'end of the α 10 cDNA, was obtained using the RACE technique. Sequence analysis of the 160 kD protein sequence showed that it was a member of the integrin α -subunit family and the protein was named α 10.

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The deduced amino acid sequence of $\alpha 10$ was found to share the general structure of the integrin α -subunits described in previously published reports (6-21). The large extracellular N-terminal part of $\alpha 10$ contains a

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seven-fold repeated sequence which was recently predicted to fold into a β -propeller domain (32). The integrin subunit $\alpha 10$ contains three putative divalent cation-binding sites (DxD/NxD/NxxxD) (53), a single spanning transmembrane domain and a short cytoplasmic domain. In contrast to most α -integrin subunits the cytoplasmic domain of α 10 does not contain the conserved sequence KxGFF (R/K) R. The predicted amino acid sequence in $\alpha 10$ is KLGFFAH. Several reports indicate that the integrin cytoplasmic 10 domains are crucial in signal transduction (54) and that membrane-proximal regions of both α - and β -integrin cytoplasmic domains are involved in modulating conformation and affinity state of integrins (55-57). It is suggested that the GFFKR motif in α -chains are important for association of integrin subunits and for transport of the integrin to the plasma membrane (58). The KxGFFKR domain has been shown to interact with the intracellular protein calreticulin (59) and interestingly, calreticulin-null embryonic stem cells are deficient in integrin-mediated cell adhesion (60). It is therefor possible that the 20 sequence KLGFFAH in all have a key function in regulating the affinity between $\alpha 10\beta 1$ and matrix proteins.

Integrin α subunits are known to share an overall identity of 20-40% (61). Sequence analysis showed that the α 10 subunit is most closely related to the I-domain containing α -subunits with the highest identity to α 1 (37%) and α 2 (35%). The integrins α 1 β 1 and α 2 β 1 are known receptors for both collagens and laminins (24;62;63) and we have also recently demonstrated that α 2 β 1 interacts with the cartilage matrix protein chondroadherin (42). Since α 10 β 1 was isolated on a collagen type II-Sepharose, we know that collagen type II is a ligand for α 10 β 1. We have also shown by affinity purification experiments that α 10 β 1 interacts with collagen type I but it remains to be seen whether laminin or chondroadherin are also ligands for this integrin.

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The $\alpha 10$ associated β -chain migrated as the $\beta 1$ integrin subunit both under reducing and non-reducing conditions. To verify that the $\alpha 10$ associated β -chain indeed is $\beta 1$, chondrocyte lysates were immunoprecipitated with antibodies against $\alpha 10$ or $\beta 1$ followed by Western blot using antibodies against the $\beta 1$ -subunit. These results clearly demonstrated that $\alpha 10$ is a member of the $\beta 1$ -integrin family. However, the possibility that $\alpha 10$ combine also with other β -chains can not be excluded.

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A polyclonal peptide antibody raised against the cytoplasmic domain of $\alpha 10$ precipitated two protein bands with M_r of approximately 160 kD (α 10) and 125 kD (β 1) under reducing conditions. Immunohistochemistry using the α 10-antibody showed staining of the chondrocytes in tissue sections of human articular cartilage. The antibody staining was clearly specific since preincubation of the antibody with the $\alpha 10$ -peptide completely abolished the staining. Immunohistochemical staining of mouse limb sections from embryonic tissue demonstrated that $\alpha 10$ is upregulated during condensation of the mesenchyme. This indicate that the integrin subunit $\alpha 10$ is important during the formation of cartilage. In 3 day old mice $\alpha 10$ was found to be the dominating collagen binding integrin subunit which point to that $\alpha 10$ has a key function in maintaining normal cartilage functions.

Expression studies on the protein and mRNA level show that the distribution of $\alpha 10$ is rather restrictive. Immunohistochemistry analyses have shown that $\alpha 10$ integrin subunit is mainly expressed in cartilage but it is also found in perichondrium, periosteum, ossification groove of Ranvier, in fascia surrounding tendon and skeletal muscle and in the tendon-like structures in the heart valves. This distribution point to that $\alpha 10$ integrin subunit is present also on fibroblasts and osteoblasts. PCR amplification of cDNA from different cell types revealed the presence of an alternatively spliced $\alpha 10$ integrin subunit. This spliced $\alpha 10$ was domi-

nating in fibroblasts which suggests that $\alpha 10$ in fibroblasts may have a different function compared to $\alpha 10$ present on chondrocytes.

Expression of the integrin subunit $\alpha 10$ was found to decrease when chondrocytes were cultured in monolayer. In contrast, the expression of $\alpha 10$ was found to increase when the cells were cultured in alginate beads. Since the latter culturing model is known to preserve the phenotype of chondrocytes the results suggest that $\alpha 10$ can function as marker for a differentiated chondrocyte.

Adhesion between tendon/ligaments and the surrounding tissue is a well-known problem after infection, injury and after surgical intervention. Adhesion between tendon and tendon sheets impairs the gliding function and cause considerable problems especially during healing of tendons in e.g. the hand and fingers leading to functional incapacity. The localisation of the α10 integrin subunit in the fascia of tendon and skeletal muscle makes α10 a possible target for drugs and molecules with antiadhesive properties that could prevent impairment of the function of tendon/ligament. The integrin subunit α10 can also be a target for drugs or molecules with antiadhesive properties in other tissues where adhesion is a problem.

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EXAMPLES

Example 1

Affinity purification of the $\alpha 10 \text{integrin}$ subunit on 30 collagen type II-Sepharose.

Materials and Methods

Bovine chondrocytes, human chondrocytes or human chondrosarcoma cells were isolated as described earlier [Holmvall et al, Exp Cell Res, 221, 496-503 (1995), Camper et al, JBC, 273, 20383-20389 (1998)]. A Triton X-100 lysate of bovine chondrocytes was applied to a fibronectin-Sepharose precolumn followed by a collagen

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type II-Sepharose column and the integrin subunit $\alpha 10$ was eluted from the collagen type II-column by EDTA (Camper et al, JBC, 273, 20383-20389 (1998). The eluted proteins were precipitated by methanol/chloroform, separated by SDS-PAGE under reducing conditions and stained with Coomassie blue. (Camper et al, JBC, 273, 20383-20389 (1998). Peptides from the $\alpha 10$ protein band were isolated by in-gel digestion with a trypsin and phase liquid chromatography and sequenced by Edman degradation (Camper et al, JBC, 273, 20383-20389 (1998). Results

Fig 1 shows EDTA-eluted proteins from the fibronectin-Sepharose (A), flow-through from the collagen type II-Sepharose column (B) and EDTA-eluted proteins from the collagen type II-Sepharose (C). The $\alpha 10$ integrin subunit 15 (160 kDa) which was specifically eluted from the collagen type II column is indicated with an arrow. Figure 2 shows the amino acid sequences of six peptides that were isolated from the bovine integrin subunit $\alpha 10$. Figures 3 a, b, and c show that the all integrin subunit is present on bovine chondrocytes (3a), human chondrocytes (3b) and human chondrosarcoma cells (3c). The affinity for collagen type II, the coprecipitation with $\beta 1$ -integrin subunit and the molecular weight of 160 kDa under reducing condi-25 tions identify the α 10 integrin subunit on the different cells. These results show that $\alpha 10$ can be isolated from chondrocytes and from chondrosarcoma cells.

Example 2

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30 Amplification of PCR fragment corresponding to bovine $\alpha 10$ integrin subunit. Materials and methods

The degenerate primers GAY AAY ACI GCI CAR AC (DNTAQT, forward) and TIA TIS WRT GRT GIG GYT (EPHHSI, reverse) were used in PCR (Camper et al, JBC, 273, 20383-20389 (1998) to amplify the nucleotide sequence corresponding to the bovine peptide 1 (Figure 2). A 900 bp

PCR-fragment was then amplified from bovine cDNA using an internal specific primer TCA GCC TAC ATT CAG TAT (SAYIQY, forward) corresponding to the cloned nucleotide sequence of peptide 1 together with the degenerate primer ICK RTC CCA RTG ICC IGG (PGHWDR, reverse) corresponding to the bovine peptide 2 (Figure2). Mixed bases were used in positions that were twofold degenerate and inosines were used in positions that are three- or fourfold degenerate. mRNA isolation and cDNA synthesis was done as earlier described (Camper et al, JBC, 273, 20383-20389 (1998)). The purified fragment was cloned, purified and sequenced as earlier described (Camper et al, JBC, 273, 20383-20389 (1998)).

Results

The nucleotide sequence of peptide 1 (Figure 2)
was obtained by PCR-amplification, cloning and sequencing of bovine cDNA. From this nucleotide sequence an
exact primer was designed and applied in PCR-amplification with degenerate primers corresponding to peptides
20 2-6 (Figure 2). Primers corresponding to peptides 1
and 2 amplified a 900 bp PCR-fragment from bovine cDNA
(Figure 4).

Example 3

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25 Cloning and sequence analysis of the human $\alpha 10$ integrin subunit

Material and methods

The cloned 900bp PCR-fragment, corresponding to bovine α 10-integrin, was digoxigenin-labelled according to the DIG DNA labelling kit (Boehringer Mannheim) and used as a probe for screening of a human articular chondrocyte λ ZapII cDNA library (provided by Michael Bayliss, The Royal Veterinary Basic Sciences, London, UK) (52). Positive clones containing the pBluescript SK+ plasmid with the cDNA insert were rescued from the ZAP vector by in vivo excision as described in the ZAP-cDNA® synthesis kit (Stratagene). Selected plasmids were purified and

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sequenced as described earlier (Camper et al, JBC, 273, 20383-20389 (1998)) using T3, T7 and internal specific primers. To obtain cDNA that encoded the 5' end of α10 we designed the primer AAC TCG TCT TCC AGT GCC ATT CGT GGG (reverse; residue 1254-1280 in α10 cDNA) and used it for rapid amplification of the cDNA 5' end (RACE) as described in the Marathon™ cDNA Amplification kit (Clontech INC., Palo Alto, CA).

Results

Two overlapping clones, hcl and hc2 (Figure 5), were 10 isolated, subcloned and sequenced. These clones contained 2/3 of the nucleotide sequence including the 3' end of the cDNA. A third clone (racel; Figure 5), which contained the 5'end of the α 10 cDNA, was obtained using the RACE technique. From these three overlapping clones of 15 alo cDNA, 3884 nucleotides were sequenced The nucleotide sequence and deduced amino acid sequence is shown in Figure 6. The sequence contains a 3504-nucleotide open reading frame that is predicted to encode a 1167 amino acid mature protein. The signal peptide cleavage site is 20 marked with an arrow, human homologues to bovine peptide sequences are underlined and the I-domain is boxed. Metal ion binding sites are indicated with a broken underline, potential N-glycosylation sites are indicated by an 25 asterisk and the putative transmembrane domain is double underlined. The normally conserved cytoplasmic sequence is indicated by a dot and dashed broken underline.

Sequence analysis demonstrate that $\alpha 10$ is a member of the integrin α -subunit family.

Example 4

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Identification of a clone containing a splice variant of $\alpha 10 \ \ :$

One clone which was isolated from the human chon- drocyte library (see Example 3) contained a sequence that was identical to the sequence of $\alpha 10$ integrin subunit except that the nucleotides between nt positions

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2942 and 3055 were deleted. The splice variant of $\alpha 10$ was verified in PCR experiment using primers flanking the splice region (see figure 14).

5 Example 5

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Identification of $\alpha 10$ integrin subunit by Northern blot

Material and methods

Bovine chondrocyte mRNA was purified using a OuickPrep®Micro mRNA Purification Kit (Pharmacia Biotech, 10 Uppsala, Sweden), separated on a 1% agarose-formaldehyde gel, transferred to nylon membranes and immobilised by UVcrosslinking. cDNA-probes were 32P-labelled with Random Primed DNA Labeling Kit (Boehringer Mannheim). Filters 15 were prehybridised for 2-4 hours at 42°C in 5x SSE, 5x Denharts solution, 0.1 % SDS, 50 μg/ml salmon sperm DNA and 50% formamide and then hybridised over night at 42 °C with the same solution containing the specific probe (0.5-1 x 106 cpm/ml). Specifically bound cDNA-20 probes were analysed using the phosphoimager system (Fuji). Filters were stripped by washing in 0.1% SDS, for 1 hour at 80° C prior to re-probing. The α 10-integrin cDNA-probe was isolated from the racel-containing plasmid using the restriction enzymes BamHI (GIBCO BRL) and NcoI (Boehringer Mannheim). The rat β 1-integrin cDNA probe was 25 a kind gift from Staffan Johansson, Uppsala, Sweden. Results

Northern blot analysis of mRNA from bovine chondrocytes showed that a human $\alpha 10$ cDNA-probe hybridised with a single mRNA of approximately 5.4 kb (Figure 7). As a comparison, a cDNA-probe corresponding to the integrin subunit $\alpha 1$ was used. This cDNA-probe hybridised a mRNA-band of approximately 3.5 kb on the same filter. These results show that a cDNA-probe against $\alpha 10$ can be used to identify the $\alpha 10$ integrin subunit on the mRNA level.

Example 6

Preparation of antibodies against the integrin subunit $\alpha 10$

A peptide corresponding to part of the all cytoplas-5 mic domain, Ckkipeeekreekle (see figure 6) was synthesised and conjugated to keyhole limpet hemocyanin (KLH). Rabbits were immunised with the peptide-KLH conjugate to generate antiserum against the integrin subunit $\alpha 10$. Antibodies recognising $\alpha 10$ were affinity purified on an peptide-coupled column (Innovagen AB).

Example 7

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Immunoprecipitation of the integrin subunit $\alpha 10$ from chondrocytes

15 Material and methods

Human chondrocytes were 125I-labelled, lyzed with Triton X-100 and immunoprecipitated as earlier described (Holmvall et al, Exp Cell Res, 221, 496-503 (1995), Camper et al, JBC, 273, 20383-20389 (1998)). Triton X-100 lysates of 125I-labeled human chondrocytes were immuno-20 precipitated with polyclonal antibodies against the integrin subunits $\beta1$, $\alpha1$, $\alpha2$, $\alpha3$ or $\alpha10$. The immunoprecipitated proteins were separated by SDS-PAGE (4-12%) under non-reducing conditions and visualised using a phospho-25 imager. Triton X-100 lysates of human chondrocytes immunoprecipitated with α10 or β1 were separated by SDS-PAGE (8%) under non-reducing conditions and analysed by Western blot using the polyclonal \$1 antibody and chemiluminescent detection as described in Camper et al, JBC, 30 273, 20383-20389 (1998).

Results

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The polyclonal peptide antibody, raised against the cytoplasmic domain of $\alpha 10$, precipitated two protein bands with Mr of approximately 160 kD (α 10) and 125 kD (β 1) under reducing conditions. The $\alpha 10$ associated β -chain migrated as the $\beta1$ integrin subunit (Figure 8a). To verify that the $\alpha 10$ associated β -chain in chondrocytes

indeed is $\beta 1$, chondrocyte lysates were immunoprecipitated with antibodies against $\alpha 10$ orb $\beta 1$ followed by Western blot using antibodies against the $\beta 1$ -subunit (Figure 8b). These results clearly demonstrated that $\alpha 10$ is a member of the $\beta 1$ -integrin family. However, the results do not exclude the possibility that $\alpha 10$ can associate with other β -chains in other situations.

Example 8

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10 Immunohistochemical staining of the integrin subunit $\alpha 10$ in human and mouse cartilage Material and methods

Frozen sections of adult cartilage (trochlear groove) obtained during surgery (provided by Anders

Lindahl, Salgrenska Hospital, Gothenburg, Sweden and frozen sections from of 3 day old mouse limb were fixed and prepared for immunohistochemistry as earlier described (Camper et al, JBC, 273, 20383-20389 (1998)). Expression of α10 integrin subunit was analysed using the polyclonal antibody against the cytoplasmic domain as a primary antibody (see Example 6) and a secondary antibody conjugated to peroxidase.

Results

Figures 9 show immunostaining of human adult articu-25 lar cartilage.

The $\alpha 10$ -antibody recognising the cytoplasmic domain of $\alpha 10$ stained the chondrocytes in tissue sections of human articular cartilage (A). The staining was depleted when the antibody was preincubated with the $\alpha 10$ - peptide (B). A control antibody recognising the $\alpha 9$ integrin subunit did not bind to the chondrocyte (C).

Figures 10 shows that the $\alpha 10$ antibody stain the majority of chondrocytes in the growing bone anlage (a and b). The $\alpha 10$ antibody also recognised cells in the ossification groove of Ranvier (b), especially the osteoblast in the bone bark which are lining the cartilage in the metaphys are highly positive for $\alpha 10$. The

PCT/SE99/00544

cells in the ossification groove of Ranvier are believed to be important for the growth in diameter of the bone. The integrin subunit $\alpha 10$ is also highly expressed in perichondrium and periosteum. Cell in these tissues are likely important in the repair of the cartilage tissue. The described localisation of the integrin subunit $\alpha 10$ suggest that this integrin is important for the function of the cartilage tissue.

10 Example 9

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Immunohistochemical staining of the integrin subunit $\alpha 10$ during mouse development $$\operatorname{\textsc{Material}}$$ and methods

Frozen sections from mouse embryos (13.5 days) were investigated for expression of $\alpha 10$ by immunhistochemistry as described in Camper et al, JBC, 273, 20383-20389 (1998). Expression of $\alpha 10$ integrin subunit was analysed using the polyclonal antibody against the cytoplasmic domain as a primary antibody (see Example 6) and a secondary antibody conjugated to peroxidase. The embryo sections were also investigated for expression of integrin subunit $\alpha 1$ (monoclonal antibody from Pharmingen) and collagen type II (monoclonal antibody, kind gift from Dr John Mo, Lund University, Sweden).

25 Results

Figure 11 show that α10 integrin subunit is unregulated in the limb when the mesenchymal cells undergo condensation to form cartilage (a). Especially the edge of the newly formed cartilage has high expression of α10.

30 The formation of cartilage is verified by the high expression of the cartilage specific collage type II (b). The control antibody against α1 integrin subunit showed only weak expression on the cartilage (c). In other experiments expression of α10 was found in all cartilage containing tissues in the 3 day old mouse including limbs, ribs and vertebrae. The upregulation of α10 during formation of cartilage suggest that this integrin subunit is

important both in the development of cartilage and bone and in the repair of damaged cartilage tissue.

Example 10

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mRNA expression of $\alpha 10$ in tissues other than articular cartilage

Material and methods

Expression of $\alpha 10$ integrin subunit was examined on the mRNA level in different human tissues. A Northern dot blot with immobilised mRNA from the listed tissues in Figure 12 was hybridised with an $\alpha 10$ integrin cDNA probe isolated from the race 1-containing plasmid using the restriction enzymes Bam H1 and Nco1. The degree of hybridisation was analysed using a phospho imager. The following symbols denote mRNA level in increasing order: -, +, ++, +++, ++++.

Results

Analysis of the hybridised mRNA showed that $\alpha 10$ was expressed in aorta, trachea, spinal cord, heart, lung, and kidney (Figure 12). All other tissues appeared negative for $\alpha 10$ expression. These results point to a restricted distribution of the $\alpha 10$ integrin subunit.

Example 11

Immunohistochemical staining of $\alpha 10$ in fascia around tendon and skeletal muscle and in tendon structures in heart valves.

Materials and methods

Frozen sections of adult cartilage (trochlear groove) obtained during surgery (provided by Anders Lindahl, Salgrenska Hospital, Gothenburg, Sweden and frozen sections from of 3 day old mouse limb were fixed and prepared for immunohistochemistry as earlier described (Camper et al, JBC, 273, 20383-20389 (1998)). Expression of α10 integrin subunit was analysed using the polyclonal antibody against the cytoplasmic domain as a pri-

mary antibody (see Example 6) and a secondary antibody conjugated to peroxidase.
Results

As shown in figures 13 expression of $\alpha 10$ was found in the fascia surrounding tendon (a) and skeletal muscle (b) and in the tendon structures in the heart valves (c). This localisation suggest that $\alpha 10$ can bind to other matrix molecules in addition to the cartilage specific collagen type II. The localisation of the integrin $\alpha 10$ on the surface of tendons indicate that $\alpha 10$ can be involved in unwanted adhesion that often occurs between tendon/ligaments and the surrounding tissue after infection, injury or after surgery.

15 Example 12

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mRNA expression of $\alpha 10$ integrin subuhit in chondrocytes, endothelial cells and fibroblasts. Material and methods

Isolation of mRNA, synthesis of cDNA and PCR ampli-20 fication was done as earlier described (Camper et al, JBC, 273, 20383-20389 (1998)). Results

Figure 14 shows PCR amplification of $\alpha 10$ cDNA from human articular chondrocytes (lanes A6 and B1), human 25 umbilical vein endothelial cells (lane A2), human fibroblasts (lane A4) and rat tendon (Fig 14b, lane B2). Lanes 1, 3, and 5 in figure:14 A show amplified fragments corresponding to the integrin subunit α2 in endothelial cells, fibroblasts and chondrocytes, respectively. cDNA-30 primers corresponding to the $\alpha 10$ sequence positions nt 2919-2943 (forward) and nt 3554-3578 (reverse) (see Figure 6) were used to amplify α10 cDNA from the different cells. The figure shows that all was amplified in all three cell types. Two fragments of $\alpha 10$ was amplified which represent the intact form of $\alpha 10$ (larger fragment) and a splice variant (smaller fragment). The larger fragment was dominating in chondrocytes while the smaller fragment was more pronounced in tendon (B2).

Example 13

Construction of $\alpha 10$ mammalian expression vector. 5 The full length protein coding sequence of all (combined from 3 clones, see figure 6) was inserted into the mammalian expression vector, pcDNA3.1/Zeo (Invitrogen). The vector contains SV40 promoter and Zeosin selection sequence. The all containing expression vector was transfected into cells that express the $\beta1$ -integrin subunit but lack expression of the all subunit. Expression of the α10 integrin subunit on the cell surface can be analysed by immunoprecipitation and/or flow cytometry using antibodies specific for $\alpha 10$. The ligand binding capacity and 15 the function of the inserted $\alpha 10$ integrin' subunit can be demonstrated in cell adhesion experiment and in signalling experiments.

20 Example 14

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Construction of mammalian expression vector containing a splice variant of $\alpha 10$.

The full length protein coding sequence of the splice variant of $\alpha 10$ (nt 2942-nt3055 deleted) was inserted into the mammalian expression vector pcDNA3 (see Example 13). Expression and function of the splice variant can be analysed as described in example 13 and compared with the intact $\alpha 10$ integrin subunit.

30 Example 15

Partial isolation and characterisation of the $\alpha 10$ integrin genomic DNA \$Material\$ and methods

Human α10 cDNA, isolated from the racel-containing plasmid using the restriction enzymes BamHI (GIBCO BRL) and NcoI (Boehringer Mannheim), was ³²P-labelled and used as a probe for screening of a mouse 129 cosmid library

(provided by Reinhard Fässler, Lund University). Positive clones were isolated and subcloned. Selected plasmids were purified and sequenced as described earlier (Camper et al, JBC, 273, 20383-20389 (1998)) using T3, T7 and internal specific primers. Primers corresponding to mouse genomic DNA were then constructed and used in PCR to amplify and identify the genomic sequence of α 10 from the cosmid clones.

Results

Figure 15 shows 7958 nt of the $\alpha 10$ gene. This partial genomic DNA sequence of $\alpha 10$ integrin contains 8 exons, and a Kozak sequence. The mouse genomic $\alpha 10$ sequence was used to generate a targeting vector for knockout experiments.

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Example 16

Human chondrocytes cultured in monolayer for 2 weeks were detached with trypsin-EDTA and introduced into alginate beads. Chondrocytes cultured in alginate are known to preserve their phenotype while chondrocytes cultured in monolayer are dedifferentiated. After 11 days chondrocytes cultured either in alginate or on monolayer were isolated and surface labelled with $^{125}\mathrm{I}$. The $\alpha10$ integrin subunit was then immunoprecipitated with polyclonal antibodies recognising the cytoplasmic domain of $\alpha10$ (see Example 6 and Camper et al, JBC, 273, 20383-20389 30 (1998)).

Results -

As shown in figure 16 chondrocytes cultured in alginate beads (lanes 3 and 4) upregulated their protein expression of $\alpha 10\beta 1$. This was in contrast to chondrocytes cultured in monolayer (lanes 1 and 2) which had a very low expression of $\alpha 10\beta 1$. Immunoprecipitation with ab control antibody is shown in lanes 1 and 3.It is known that

chondrocytes preserve their cartilage specific matrixproduction in alginate cultures but not in monolayer culture which point to that alginate preserve the phenotype of chondrocytes. These results support that $\alpha 10$ integrin subunit can be used as a marker for differentiated chondrocytes.

Example 17

 $\label{eq:continuous} Immunoprecipitation of the $\alpha 10$ integrin subunit from 10 human smooth muscle cells.$

Material and methods

Human smooth muscle cells were isolated from human aorta. After one week in culture the cells were ¹²⁵I-labelled, lysed and immunoprecipitated with antibodies against the integrin subunit β1 (lane 1), α1 (lane 2), α2 (lane 3), α10 (lane 4), α3 (lane 5), control (lane 6) (Figure 17). The experiment was done as described in Example 7.

Results

The $\alpha 10$ antibody precipitated two bands from the smooth muscle cells corresponding to the $\alpha 10$ and the $\beta 1$ integrin subunit (Fig. 17).

Example 18

25 Construction of bacterial expression vector containing sequence for $\alpha 10$ splice region.

A plasmid for intracellular expression in E. coli of the alternatively spliced region (amino acid pos. 952-986, SEQ. ID 1) was constructed as described. The alternatively spliced region were back-translated using the E. coli high frequency codon table, creating a cDNA sequence of 96% identity with the original sequence (SEQ. ID 1 nucleotide pos 2940-3044). Using sequence overlap extension (Horton et al., Biotechniques 8:528, 1990) primer α10pfor (tab. I) and α10prev (tab. I) was used to generate a double stranded fragment encoding the α10 amino acid sequence. This fragment was used as a PCR

template with primers $\alpha 10 pfor 2$ (tab. I) and $\alpha 10 prev 2$ (tab. I) in order to generate restriction enzyme site for sub-cloning in a pET vector containing the Z-domain of staphylococcal protein A, creating a fusion of the $\alpha 10$ spliced region with the amino terminal of the Z-domain with trombin cleavage site residing in-between. The fragment generated in the second PCR reaction is shown (SEQ ID No. 3) also indicating the unique restriction enzymes used for sub-cloning in the expression vector.

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Table I

α10pfor	5'- GTTCAGAACCTGGGTTGCTACGTTGTTTCCGGTCTGATCATCTCCGC TCTGCTGCCGGCTGT-3'					
α10pfor2	5'-GGGGCATATGGTTCAGAACCTGGGTTGCTACGTTG-3'					
α10prev	5'- GATAACCTGGGACAAGCTTAGGAAGTAGTTACCACCGTGAGCAACAG CCGGCAGCAGAGCGGA-3'					
α10prev2	5'- GGGGGGATCCGCGCGCACCAGGCCGCTGATAACCTGGGACAAGCTT AGGAAGT-3'					

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• •	ENERAL INFORMATION: NUMBER OF SEQUENCES: 2	
(i) (7 (1 (0	NFORMATION FOR SEQ ID NO. 1: SEQUENCE CHARACTERISTICS: A) LENGTH: 3884 base pairs B) TYPE: nucleic acid and amino acid C) STRANDEDNESS: double D) TOPOLOGY: linear	
(vi) (1 (1	MOLECULAR TYPE: cDNA ORIGINAL SOURCE: E) ORGANISM: human F) CELLTYPE: chondrocyte) SEQUENCE DESCRIPTION: SEQ ID NO. 1:	
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	2881	GTTCA	Q CCC3	Y ATA:	E I'GG	P GAC	H CCT	L	L AGT	F GGG	SAG	S FGG(E CCC	S AGAI	T T	L CÁA!	H AAC	R CAC	GAT! Y ICTO	ACTÇ E CAGG	2880
•	2881	GTTCA CAAGT	Q ACCCI ACCCI	Y ATAT -+	E TGGG ACCG	P GACO	H CCT(+- GGA(L CCC GGG	L AGT ICA	F GGG'	SAG S TCC AGG	S TGG(E CCCI	S AGAZ -+	T T ATTO	L CÁA/ STT	H AACO TTGO	R CACT	Y Y FCT(AGA(E CAGG + GTCC	- 2940
a	2881	GTTCA CAAGI V H	Q ACCCI GGGT	Y ATAT -+ PATA Y	E TGGG ACCG	P GACO CTGO T	H CCTC SGAC	L CCC GGG	L AGT ICA	F GGG CCC	S FCC + AGG	S TGG(ACC(E CCCI GGG1	S AGA/ -+ FCT? E	T ATTO TAAO F	L CÁA! STT:	H AACO TTGO T	R CAC GTG/	Y TCTO AGAO	E CAGG + GTCC R	- 2940
a		GTTCA CAAGT V H	Q CCC? CGGGT P	Y ATAT TAT Y TGC	E TGGG ACCG G AAGG	P GACO CTGO T	H CCTC GGAC L	L CCC GGG P AGT	L AGT(ICA) V	E GGG' G G GAA	SAGA STCC HAGGA PCCTC	S TGGC ACCC	E CCCA GGG1 P	S AGAI FCTT E ACCO	T ATTO TAAO F	L CÁA! STT: K	H AACO TTGO T	R CACT STGA T	Y FCT(AGA(L	E CAGG GTCC R GCAT	- 2940 -
	2941	GTTCA CAAGT V H ACTAA TGATT	Q ACCCA P ACAAT	Y ATAT TATA Y TGCA ACGT	E FGGG ACCC G AAGC	P GACO T CTGO GACO	H CCTC GGAC L CATA	L CCC GGC P AGT CA	L AGTO FCAO V SCAO	F SGG CCC G G SAA	SAGA	S TGGGGACCC	E CCCA	S AGAI FCTT E ACCO	T ATTO	L CAA! STTT K AGGG	H AACO TTGO T CCCA	R CACC	Y ICTO AGA L IGTO	E CAGG+ STCC R GCAT+ CGTA	- 2940 -
a	2941	GTTCA CAAGT V H ACTAA TGATT	Q ACCCA P ACAA ACAA ACAA ACAA N	Y ATATATATATATATATATATATATATATATATATATAT	E TGGGG G AAGCC	P GACO T CTGO GACO C	H CCTO	L CCCA GGGG P AGTO V	L AGTO V V GCAO CGTO	F GGG G G GGAA CTTC	SAGA S FCCT P FCCT SGA	S S TGGGGGACCC	E E E E E E E E E E E E E E E E E E E	S AGAI TCTT E ACCC	T TATTO	L CAAAA K K AAGGG	H AACO TT CCCCA	R CACC	Y ICTO AGAGA L IGTO	E CAGG+ STCC R SCAT+ CGTA	- 2940 -
a	2941	GTTCA CAAGT V H ACTAA TGATT T N CCAGA	Q ACCCI P ACAAT N AGGAA	Y ATATATATATATATATAACGT A GCTT	E TGGGG G G S TTTCG	P GACO T CTGO C GACO	H CCT(CTC) L CATA THE CATA THE CACA	L CCCA FP PAGTO V V	L AGT(F GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	SAGGE	S TGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	E CCCA	S AGAI	T ATTO	L CAAI K K AGGG CCC G	H AACC TTGC T T CCCCA P STGT	R CACC	Y FCT(AGA(L FGT(ACA(V	E CAGG+ STCC R SCAT+ CGTA H GGTG	- 2940 - 3000
а	2941	GTTCA CAAGT V H ACTAA TGATT T N CCAGA	Q ACCCI P ACCAAT N AGGAAC CCCTC	Y ATATA TATA Y TGCA ACGT A GCTT CGGA	E FGGG G AAGGC TTTCG S FCAA	P GACO T CTGO GACAO C GRETO	H CCTC GGAC L CATA THE CACA TH	L CCCA P AGT(V AAAA	L AGTO V GCAC CGTO Q CAG	EAAACTTGA	SAGA	S IIGGC G T IIGGC ACCC	E CCCA	S AGAM FCTT E ACCO P CAAT	T ATTO	L CAAAA K K AAGGO G CCAO	H AACCAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAA	R CACT	Y FCT(AGA(L FGT(ACA(V FGT(CCCA(ACTC E CAGG+ GGTCC R GCAT+ CGTA H GGTG	- 2940 - 3000 - 3060
a	2941	GTTCA CAAGT V H ACTAA TGATT T N CCAGA	Q ACCCI P ACAAT N AGGAC CCCCC	Y ATATATATATATATATATATATATATATATATATATAT	E FGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	P GACO T CTGO CACAC ACAC H	H CCT(GGAC L CATA I CAC T	L CCCA	L AGTO V GCAG	F GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	S S TCC' AGG. P CCTC' GGAA' L CTTT	S IIGGO G G G G G G G G G G G G G G G G G	E CCCA P FGAACTT	S AGAI F ACCO P CAAT N	T TANGE F P PACT	L CAAI	H AACC	R CACT T ACCT P CCAC	Y TCT(AGA(L TGT(ACA(V TGT(CCA(V	ACTC E CAGG CAGG R GCAT CGTA H GGTG V	- 2940 - 3000 - 3060

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	3601		PCT	GTT															AGG		+ GAGG	3660								
	:	TCGTGATCCTGGTTCCATAGCCAACACTGGGGCTTTTGTTTG																												
	3661		CAC	TAG	GAC	CAA	GGT	ATC	GGT	rgt	GAC	CCC		AAC	-			-		AGG	+ 3720 GGGTC									
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- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 143 base pairs
 - (B) TYPE: nucleic acid and amino acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
- (iii) MOLECULAR TYPE: cDNA
- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: human
 - (B) CELLTYPE: chondrocyte
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO. 3:

NdeI

GGGGCATATGGTTCAGAACCTGGGTTGCTACGTTGTTTCCGGTCTGATCATCTCCGCTCT

1 -----+ 60
CCCCGTATACCAAGTCTTGGACCCAACGATGCAACAAAGGCCAGACTAGTAGAGGCGAGA

b - G H M V Q N L G C Y V V S G L (I I S A L -

GCTGCCGGCTGTTGCTCACGGTGGTAACTACTTCCTAAGCTTGTCCCAGGTTATCAGCGG
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CGACGGCCGACAACGAGTGCCACCATTGATGAAGGATTCGAACAGGGTCCAATAGTCGCC

b LPAVAHGGNYFLSLSQVISG-

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b LVPRGSP -

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CLAIMS

- 1. A recombinant or isolated integrin subunit $\alpha 10$ comprising the amino acid sequence shown in SEQ ID No. 1 or SEQ ID No. 2, or homologues or fragments thereof having similar biological activity.
- 2. A process of producing a recombinant integrin subunit $\alpha 10$ comprising the amino acid sequence shown in SEQ ID No. 1 or SEQ ID No. 2, or homologues or fragments thereof having similar biological activity, which process comprises the steps of
- a) isolating a polynucleotide comprising a nucleotide sequence coding for an integrin subunit $\alpha 10$, or homologues or fragments thereof having similar biological activity,
 - b) constructing an expression vector comprising the isolated polynucleotide,
- c) transforming a host cell with said expressionvector,
 - d) culturing said transformed host cell in a culture medium under conditions suitable for expression of integrin subunit $\alpha 10$, or homologues or fragments thereof having similar biological activity, in said transformed host cell, and, optionally,
 - e) isolating the integrin subunit $\alpha 10$, or homologues or fragments thereof having similar biological activity, from said transformed host cell or said culture medium.
 - 3. A process of providing an integrin subunit α10, or homologues or fragments thereof having similar biological activity, whereby said subunit is isolated from a cell in which it is naturally present.
 - 4. An isolated polynucleotide comprising a nucleotide coding for an integrin subunit $\alpha 10$, or for homologues or fragments thereof, which polynucleotide comprises the nucleotide sequence shown in SEQ ID No. 1 or SEQ ID No. 2 or suitable parts thereof.

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- 5. An isolated polynucleotide or oligonucleotide which hybridises to a DNA or RNA encoding an integrin subunit $\alpha 10$ or homologues or fragments thereof, wherein said polynucleotide or oligonucleotide fails to hybridise to a DNA or RNA encoding an integrin subunit $\alpha 1$.
- 6. A vector comprising a polynucleotide or oligonucleotide coding for an integrin subunit $\alpha 10$, or homologues or fragments thereof, which polynucleotide or oligonucleotide comprises the nucleotide sequence shown in SEQ ID No. 1 or SEQ ID No. 2 or parts thereof.
- 7. A vector comprising a polynucleotide or oligonucleotide which hybridises to a DNA or RNA encoding an integrin subunit α10 or homologues or fragments thereof,
 15 wherein said polynucleotide or oligonucleotide fails to hybridise to a DNA or RNA encoding an integrin subunit α1.
 - 8. A cell containing the vector as defined in any one of claims 6 and 7.
- 9. A cell generated by the process in claim 2, in which a polynucleotide or oligonucleotide coding for an integrin subunit α10, or homologues or fragments thereof, which polynucleotide or oligonucleotide comprises the nucleotide sequence shown in SEQ ID No. 1 or SEQ ID No. 2 or parts thereof has been stably integrated in the cell genome.
 - 10. Binding entities having the capability of binding specifically to integrin subunit $\alpha 10$ comprising the amino acid sequence of SEQ ID No. 1 or SEQ ID No. 2, or to homologues or fragments thereof.
 - 11. Binding entities according to claim 10, which are chosen from the group comprising proteins, peptides, carbohydrates, lipids, natural integrin binding ligands, polyclonal and monoclonal antibodies, and fragments thereof.
 - 12. A recombinant or isolated integrin heterodimer comprising a subunit $\alpha 10$ and a subunit β , in which the

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subunit $\alpha 10$ comprises the amino acid sequence shown in SEQ ID No. 1 or SEQ ID No. 2, and homologues and fragments thereof having similar biological activity.

- 13. A recombinant or isolated integrin heterodimer according to claim 12, wherein the subunit β is β 1.
- 14. A process of producing a recombinant integrin heterodimer comprising a subunit $\alpha 10$ and a subunit β , in which the subunit $\alpha 10$ comprises the amino acid sequence shown in SEQ ID No. 1 or SEQ ID No. 2, and homologues and fragments thereof, which process comprises the steps of
- a) isolating one polynucleotide comprising a nucleotide sequence coding for a subunit $\alpha 10$ of an integrin heterodimer and, optionally, another polynucleotide comprising a nucleotide sequence coding for a subunit β of an integrin heterodimer, or polynucleotides or oligonucleotides coding for homologues or fragments thereof having similar biological activity,
- b) constructing an expression vector comprising said isolated polynucleotide coding for said subunit $\alpha 10$ optionally in combination with an expression vector comprising said isolated nucleotide coding for said subunit β ,
- c) transforming a host cell with said expression vector or vectors,
- d) culturing said transformed host cell in a culture medium under conditions suitable for expression of an integrin heterodimer comprising a subunit $\alpha 10$ and a subunit β , or homologues or fragments thereof having similar biological activity, in said transformed host cell, and, optionally,
- e) isolating the integrin heterodimer comprising a subunit $\alpha 10$ and a subunit β , or homologues or fragments thereof having similar biological activity, or the $\alpha 10$ subunit thereof from said transformed host cell or said culture medium.
- 15. A process of providing an integrin heterodimer comprising a subunit $\alpha 10$ and a subunit $\beta\text{,}$ or homologues

or fragments thereof having similar biological activity, whereby said integrin heterodimer is isolated from a cell in which it is naturally present.

- 16. A cell containing a first vector, said first vector comprising a polynucleotide or oligonucleotide coding for a subunit $\alpha 10$ of an integrin heterodimer, or for homologues or parts thereof having similar biological activity, which polynucleotide or oligonucleotide comprises the nucleotide sequence shown in SEQ ID No. 1 or SEQ ID No. 2 or parts thereof, and a second vector, said second vector comprising a polynucleotide or oligonucleotide coding for a subunit β of an integrin heterodimer, or for homologues or fragments thereof.
- 17. Binding entities having the capability of binding specifically to the integrin heterodimer comprising a subunit $\alpha 10$ and a subunit β , or to homologues or fragments thereof, or a subunit $\alpha 10$ thereof, having similar biological activity.
 - 18. Binding entities according to claim 17, wherein the subunit β is β 1.
 - 19. Binding entities according to claim 17 or 18, which are chosen among the group comprising proteins, peptides, carbohydrates, lipids, natural integrin binding ligands, and fragments thereof.
- 20. A fragment of the integrin subunit $\alpha 10$, which fragment is a peptide chosen from the group comprising peptides of the cytoplasmic domain, the I-domain and the spliced domain.
- 21. A fragment according to claim 20, which is a peptide comprising the amino acid sequence KLGFFAHKKIPEEEKREEKLEQ.
 - 22. A fragment according to claim 20, which comprises the amino acid sequence from about amino acid No. 952 to about amino acid no. 986 of SEQ ID No. 1.
- 35 23. A fragment according to claim 20, which is a peptide comprising the amino acid sequence from about

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amino acid No. 140 to about amino acid no. 337 of SEO ID No. 1.

- 24. A method of producing a fragment of the integrin subunit $\alpha 10$ as defined in any one of claims 20-23, which method comprises a sequential addition of amino acids containing protective groups.
- 25. A polynucleotide or oligonucleotide coding for a fragment of the integrin subunit $\alpha 10$ as defined in any one of claims 20-23.
- 10 26. Binding entities having the capability of binding specifically to a fragment of the human integrin subunit $\alpha 10$ as defined in any one of claims 20-23.
 - 27. Binding entities according to claim 26, which are chosen from the group comprising proteins, peptides, carbohydrates, lipids, natural integrin binding ligands, and fragments thereof.
 - 28. A process of using an integrin subunit $\alpha 10$ comprising the amino acid sequence shown in SEQ ID No. 1 or SEQ ID No. 2, or an integrin heterodimer comprising said subunit $\alpha 10$ and a subunit β , or a homologue or fragment of said integrin or subunit having similar biological activity, as a marker or target molecule of cells or tissues expressing said integrin subunit $\alpha 10$, which cells or tissues are of animal including human origin.
 - 29. A process according to claim 28, whereby said fragment is a peptide chosen from the group comprising peptides of the cytoplasmic domain, the I-domain and the spliced domain.
- 30. A process according to claim 29, whereby said fragment is a peptide comprising the amino acid sequence KLGFFAHKKIPEEEKREEKLEQ.
- 31. A process according to claim 29, whereby said fragment comprises the amino acid sequence from about amino acid no. 952 to about amino acid no. 986 of SEQ ID No. 1.
 - 32. A process according to claim 29, whereby said fragment comprises the amino acid sequence from about

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amino acid no. 140 to about amino acid no. 337 of SEQ ID No. 1.

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- 33. A process according to claim 28, whereby the subunit β is $\beta1$.
- 34. A process according to claim 28, whereby said cells are chosen from the group comprising chondrocytes, smooth muscle cells, endothelial cells, osteoblasts and fibroblasts.
- 35. A process according to any one of claims 28-34, which process is used during pathological conditions involving said subunit $\alpha 10$.
 - 36. A process according to claim 35, which pathological conditions comprise damage of cartilage.
- 37. A process according to claim 36, which patho15 logical conditions comprise trauma, rheumatoid arthritis
 and osteoarthritis.
 - 38. A process according to any one of claims 28-34, which is a process for detecting the formation of cartilage during embryonal development.
- 39. A process according to any one of claims 28-34, which is a process for detecting physiological or therapeutic reparation of cartilage.
 - 40. A process according to any one of claims 28-34, which is a process for selection and analysis, or for sorting, isolating or purification of chondrocytes.
 - 41. A process according to any one of claims 28-34, which is a process for detecting regeneration of cartilage or chondrocytes during transplantation of cartilage or chondrocytes.
 - 42. A process according to any one of claims 28-34, which is a process for in vitro studies of differentiation of chondrocytes.
 - 43. A process of using binding entities having the capability of binding specifically to an integrin subunit $\alpha 10$ comprising the amino acid sequence shown in SEQ ID No. 1 or SEQ ID No. 2, or an integrin heterodimer comprising said subunit $\alpha 10$ and a subunit β , or to homo-

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logues or fragments thereof having similar biological activity, as markers or target molecules of cells or tissues expressing said integrin subunit $\alpha 10$, which cells or tissues are of animal including human origin.

- 44. A process according to claim 43, whereby said fragment is a peptide chosen from the group comprising peptides of the cytoplasmic domain, the I-domain and the spliced domain.
- 45. A process according to claim 43, whereby said
 10 fragment is a peptide comprising the amino acid sequence
 KLGFFAHKKIPEEEKREEKLEQ.
 - 46. A process according to claim 43, whereby said fragment comprises the amino acid sequence from about amino acid no. 952 to about amino acid no. 986 of SEQ ID No. 1.
 - 47. A process according to claim 43, whereby said fragment comprises the amino acid sequence from about amino acid no. 140 to about amino acid No. 337 of SEQ ID No. 1.
 - 48. A process according to claim 43, whereby the subunit β is $\beta1$.
 - 49. A process according to any one of claims 43-48, which is a process for detecting the presence of an integrin subunit $\alpha 10$ comprising the amino acid sequence shown in SEQ ID No. 1 or SEQ ID No. 2, or of an integrin heterodimer comprising said subunit $\alpha 10$ and a subunit β , or of homologues or fragments thereof having similar biological activity.
 - 50. A process according to any one of claims 43-48, which process is a process for determining the differentiation-state of cells during embryonic development, angiogenesis, or development of cancer.
 - 51. A process for detecting the presence of an integrin subunit $\alpha 10$, or of a homologue or fragment of said integrin subunit having similar biological activity, on cells, whereby a polynucleotide or oligonucleotide chosen from the group comprising a polynucleotide or oligo-

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nucleotide shown in SEQ ID No. 1 is used as a marker under hybridisation conditions wherein said polynucleotide or oligonucleotide fails to hybridise to a DNA or RNA encoding an integrin subunit $\alpha 1$.

- 52. A process according to claim 51, whereby said cells are chosen from the group comprising chondrocytes, smooth muscle cells, endothelial cells, osteoblasts and fibroblasts.
- 53. A process according to claim 51, whereby said fragment is a peptide chosen from the group comprising peptides of the cytoplasmic domain, the I-domain and the spliced domain.
- 54. A process according to claim 53, whereby said fragment is a peptide comprising the amino acid sequence KLGFFAHKKIPEEEKREEKLEQ.
- 55. A process according to claim 53, whereby said fragment comprises the amino acid sequence from about amino acid No. 952 to about amino acid no. 986 of SEQ ID No. 1.
- 56. A process according to claim 53, whereby said fragment comprises the amino acid sequence from about amino acid No. 140 to about amino acid No. 337 of SEQ ID No. 1.
 - 57. A process according to any one of claims 43-48, which is a process for determining the differentiation-state of cells during development, in pathological conditions, in tissue regeneration or in therapeutic and physiological reparation of cartilage.
- 58. A process according to claim 57, wherein the 30 pathological conditions are any pathological conditions involving the integrin subunit $\alpha 10$.
 - 59. A process according to claim 58, whereby said pathological conditions are rheumatoid arthritis, osteo-arthrosis or cancer.
- 35 60. A process according to claim 57, whereby said cells are chosen from the group comprising chondrocytes,

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smooth muscle cells, endothelial cells, osteoblasts and fibroblasts.

- 61. A process for determining the differentiation-state of cells during development, in pathological conditions, in tissue regeneration and in therapeutic and physiological reparation of cartilage, whereby a polynucleotide or oligonucleotide chosen from the nucleotide sequence shown in SEQ ID No. 1 is used as a marker under hybridisation conditions wherein said polynucleotide or oligonucleotide fails to hybridise to a DNA or RNA encoding an integrin subunit α 1.
- 62. A process according to claim 61, whereby said polynucleotide or oligonucleotide is a polynucleotide or oligonucleotide coding for a peptide chosen from the group comprising peptides of the cytoplasmic domain, the I-domain and the spliced domain.
- 63. A process according to claim 62, whereby said polynucleotide or oligonucleotide is a polynucleotide or oligonucleotide coding for a peptide comprising the amino acid sequence KLGFFAHKKIPEEEKREEKLEQ.
- 64. A process according to claim 62, whereby said peptide comprises the amino acid sequence from about amino acid no. 952 to about amino acid no. 986 of SEQ ID No. 1.
- 25 65. A process according to claim 62, whereby said peptide comprises the amino acid sequence from about amino acid no. 140 to about amino acid no. 337 of SEO ID No. 1.
 - 66. A process according to claim 61, whereby said pathological conditions are any pathological conditions involving the integrin subunit $\alpha 10$.
 - 67. A process according to claim 66, whereby said pathological conditions are rheumatoid arthritis, osteo-arthrosis or cancer.
- 35 68. A process according to claim 66, whereby said pathological conditions are atherosclerosis or inflammation.

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- 69. A process according to any one of claims 61-68, whereby said cells are chosen from the group comprising chondrocytes, smooth muscle cells, endothelial cells, osteoblasts and fibroblasts.
- 70. A pharmaceutical composition comprising as an active ingredient a pharmaceutical agent or an antibody which is capable of using an integrin heterodimer comprising a subunit $\alpha 10$ and a subunit β , or the subunit $\alpha 10$ thereof, or a homologue or fragment of said integrin or subunit $\alpha 10$ having similar biological activity, as a target molecule.
- 71. A pharmaceutical composition according to claim 70, for use in stimulating, inhibiting or blocking the formation of cartilage, bone or blood vessels.
- 72. A pharmaceutical composition according to claim 70, for use in preventing adhesion between tendon/ligaments and the surrounding tissue after infection, inflammation and after surgical intervention where adhesion impairs the function of the tissue.
- 73. A vaccine comprising as an active ingredient an integrin heterodimer comprising a subunit $\alpha 10$ and a subunit β , or the subunit $\alpha 10$ thereof, or a homologue or fragment of said integrin or subunit $\alpha 10$, or DNA or RNA coding for said integrin subunit $\alpha 10$.
 - 74. Use of the integrin subunit $\alpha 10$ as a marker or target in transplantation of cartilage or chondrocytes.
 - 75. A method of using binding entities having the capability of binding specifically to an integrin subunit $\alpha 10$ comprising the amino acid sequence shown in SEQ ID No. 1 or SEQ ID No. 2, or an integrin heterodimer comprising said subunit $\alpha 10$ and a subunit β , or to homologues or fragments thereof having similar biological activity, for promoting adhesion of chondrocytes and/or osteoblasts to surfaces of implants to stimulate osseointegration.
 - 76. Use of an integrin heterodimer comprising an integrin subunit $\alpha 10$ and a subunit β , or the subunit $\alpha 10$

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thereof, or a homologue or fragment of said integrin or subunit all having similar biological activity, as a target for anti-adhesive drugs or molecules in tendon, ligament, skeletal muscle or other tissues where adhesion impairs the function of the tissue.

- 77. A method of stimulating, inhibiting or blocking the formation of cartilage or bone, comprising administration to a subject a suitable amount of a pharmaceutical agent or an antibody which is capable of using an integrin heterodimer comprising a subunit $\alpha 10$ and a subunit β , or the subunit $\alpha 10$ thereof, or a homologue or fragment of said integrin or subunit all having similar biological activity, as a target molecule.
- 78. A method of preventing adhesion between tendon/ ligaments and the surrounding tissue after infection, inflammation and after surgical intervention where adhesion impairs the function of the tissue, comprising administration to a subject a suitable amount of a pharmaceutical agent or an antibody which is capable of using a 20 integrin heterodimer comprising a subunit α 10 and a subunit β , or the subunit $\alpha 10$ thereof, or a homologue or fragment of said integrin or subunit all having similar biological activity, as a target molecule.
 - 79. A method of stimulating extracellular matrix synthesis and repair by activation or blockage of an integrin heterodimer comprising a subunit lpha 10 and a subunit β , or of the subunit $\alpha 10$ thereof, or of a homologue or fragment of said integrin or subunit lpha 10 having similar biological activity.
- 80. A method of in vitro detecting the presence of 30 integrin binding entities, comprising interaction of an integrin heterodimer comprising a subunit $\alpha 10$ and a subunit β , or the subunit $\alpha 10$ thereof, or a homologue or fragment of said integrin or subunit, with a sample, thereby causing said integrin, subunit $\alpha 10$, or homologue 35 or fragment thereof having similar biological activity,

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to modulate the binding to its natural ligand or other integrin binding proteins present in said sample.

- 81. A method of in vitro studying consequences of the interaction of a human heterodimer integrin comprising a subunit $\alpha 10$ and a subunit β , or the subunit $\alpha 10$ thereof, or a homologue or fragment of said integrin or subunit, with an integrin binding entity and thereby initiate a cellular reaction.
- 82. A method according to claim 81, whereby the con10 sequences of said interactions are measured as alterations in cellular functions.
 - 83. A method of using DNA or RNA encoding an integrin subunit $\alpha 10$ or homologues or fragments thereof as a target molecule.
 - 84. A method according to claim 83, whereby a polynucleotide or oligonucleotide hybridises to the DNA or RNA encoding an integrin subunit $\alpha 10$ or homologues or fragments thereof and whereby said polynucleotide or oligonucleotide fails to hybridise to a DNA or RNA encoding en integrin subunit $\alpha 1$.
 - 85. A method of using a human heterodimer integrin comprising a subunit $\alpha 10$ and a subunit β , or the subunit $\alpha 10$ thereof, or a homologue or fragment of said integrin or subunit, or a DNA or RNA encoding an integrin subunit $\alpha 10$ or homologues or fragments thereof, as a marker or target molecule during angiogenesis.
 - 86. A pharmaceutical composition comprising as an active ingredient a pharmaceutical agent or an antibody which is capable of stimulating cell surface expression of an integrin heterodimer comprising a subunit $\alpha 10$ and a subunit β , or the subunit $\alpha 10$ thereof, or a homologue or fragment of said integrin or subunit $\alpha 10$ having similar biological activity.

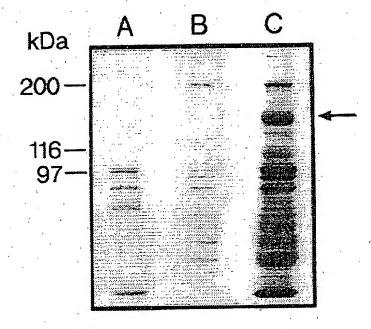
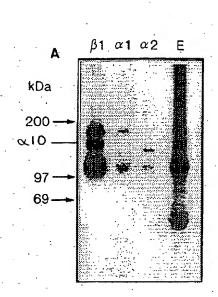
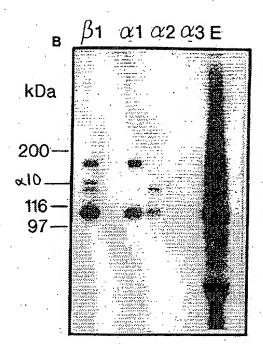


FIGURE 1

	Peptide	Amino acid sequence						
•								
	1	DNTAQTSAYIQYEPHHSI						
	2	GPGHWDR						
	3	AAFDGSGQR						
	4	FAMGALPD						
	5	FTASLDEWTTAAR						
	6.	VDASFRPQGXLAP						

FIGURE 2





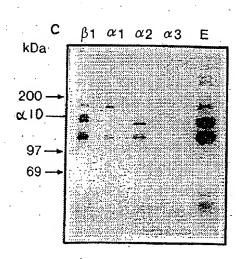


FIGURE 3

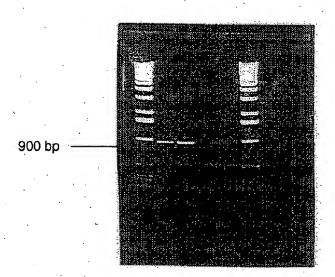


FIGURE 4

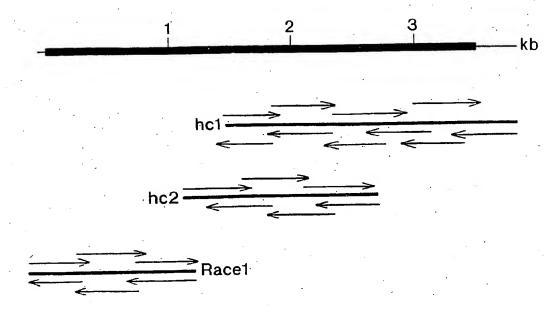


FIGURE 5

caggicagaaacceateaggacteccactegetracteactegitritecectegicytecteaca $\tt M$ $\tt E$ $\tt L$ $\tt P$ $\tt P$ $\tt V$ $\tt Y$ $\tt H$ $\tt L$ $\tt P$ $\tt L$ $\tt P$ $\tt L$ $\tt Y$ $\tt P$ $\tt L$ $\tt Y$	72 -6 .	CATCUTGCUCAGAGGATTGCTGCCTCCATGCCACATGCCCTCAGCTACTTTGGCCGAAGTGTGGATGGT H P A O R I A A A S M P H A L S Y F G R S V D G	1877 395
COTOTOTOTOTOTOTOTOTOTOTOTOTOTOTOTOTOTOT	144° 19	COSCINGATCIOCATOCACATEATCICACTCACTCACTCACCCACCCACCCACCCATCCTCCTCACC	1944 619
GGATACAGTGTCTTACAACATGTTGGGGGTGGGACAGCGATGGATG	216 41	**CCCCCCCATTGTCCATCTCACCCCATCACTGCGCCCCCCACGCCATCCGTGTGCACGCGCCC S R P 1 V H L T P S L E V T P Q A 1 S V V Q R D	2016 643
TCACCCCACCGCACCGCACCGTTATCCCCCCTGTACCGCCCCACACACA	261 67	TGTAGGGGGGGGAAGAAGAGGGTGTGTGTGGACCCCTTTGCTTCCAAGTGACCTGCGTACTCCT	2088 667
CACTIAGGIGACTACCACTOGGLANTICATCTCATCCCCCTGTGAATATCCACCTGGGGATGTCTCTGTTA H L G D T Q L G N S S N P A V N H H L G H S L L	360 91	. GCTCGCTGGCATCACCAATICTACATGAGGTCAEGGCATGACGATGATGGATGGATGGCGGAAGGTGCAGGTGCAGGTGCAGGTGATGATGGATG	2106
CACACACATOCTCATCCCCCATTCATCCCCTCTCTCCCCCTCTCTCCTCTCTCTCTTCT	432 115	CCATTIGATOGCTCTGGGCAGAGTTGTCCCCTCGGGCGGCTCAGTCTGGGGAATGTCATTGTGAG	2212 715
MITRITICACEATATOTOCCCGTGGATGCTTCATTCCAGCCTCAGGGAAGCCTGGCACCCCCACGCCCACCCCCACGCCCACGCCCACGCCCACGCCCACGCCCCACGCCCCACGCCCCACGCCCACGCCCCACGCCCCACGCCCCACGCCCCACGCCCCACGCCCCACGCCCCACGCCCCACGCCCCACGCCCCACGCCCCACGCCCCACGCCCCACGCCCCACGCCCCACGCCCCACGCCCCACGCCCCACGCCCACGCCCCACGCCCCACGCCCCACGCCCCACGCCCCACGCCCCACGCCCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCACGCCCACGCCCACGCCCACGCCCACGCCACGCCCACGCCCACGCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCCACGCCACGCCCACGCCCACGCCCACGCCCACGCCACGCCCACGCCACGCCACGCCACGCCACGCCACGCCACGCCACGCCCACGCCACGCCACGCCACGCCACGCCACGCCACGCCCACGCACGCACGCACGCCACGACG	\$04 139	CARCTACACTICCATGRACTACATCACATTACCTCCGCCCCAGTGCCCTTGACTGTGCCTTTCCCTTC O L H F H V L O T 'S D Y L R P V A L T V T F A L	2304
TOCCCAMENTACATIGATETICATICATICATICATICATICAL TICAMENICATICACIC TOCCCTOCATICAMETICAE C P T Y N . D V V I V L D G S N S I Y P W S E V . O	576 . 161	GACANTACTACANGCCAGGGCCTGTGCTGTATGAGGGCTCACCCACCCCTATACAAAAGCTGGTCCCCTTC D H T T K P G P V L N E G S P T S I C K L V F T	2376
ACCITCCTACGAACACTGGTAGGCAAACTGTTTATTGACCCAGAACAGATACAGTGGGGCCTGGTACAGTAT T F L R R L V G K L F 1 D P E Q 1 Q V G L V Q Y	643 187	TEANAGENTICIGECCTEACANTEANISTSTEACAGACCTECTCENCTCANGTCANTATGACANTCAGCCC S K D C G P D N E C V T D L V L Q V N N D I R G	2448 787
GECCAGACCOCTGTACATGAGTGCTCCCTGGGAGATTTCCCGAACGAAGGAGTAGTACTGGGAGACAACGAAG G E S P V H B W S L G D F R T K B E V V R A A K	720 211	TECAGGAGGECCECATTTCTGGTTCGAGGTGGCCGGGGGAAGTGCTGGTATCTACAGCTCTGGAGAGAGA	2520 811
AMCCICACTCGGCGGGGGGGGGAACAAAGACTGCCCAAGAAAATAATGGTGGGCTGCACGAAGAGGTTC R L S R R E G R E T R T A O A I H V A C T E G f	792 235	ANGGAMATGETTACATACGAGCETGAGTATCATCTTCTCTAGAAACCTCCACCTGGCCAGTCTCACTCCT K & N A Y N T S L S I I F S R N L II L A S L T P	2592 035
AGTCAGTCCCATGGGGCCGAGCCGAGGCTGCCAGGGCTACTGGTGGTGACTGATGGAGAGTCCCATGAT S 0 S H G G R P E A A R L L V V V T D G E S H D	861 259	CAGNENGAGCCCCTATANAGGTEGATGTCCCCCCCCTTCTCCTCATCCCCCCCCTCTCCAGGTGTCCCCCATC	2664 859
GENGAGGACCTICCTOCAGCACTAAAGOCCTGGAAGACTGACAACGACTACGGATTGCAGTCCTT G E E L P A A L K A C E A G R V T R Y G I A V L	936 203	COTGRETTCAGACTGAGCCAAGGTGACCTTTCTGCTAGAGTTTAGGTTTAGCTGCTGCTCCTCTCCTCGCGACC F V F Q T G A K V T F L L E F E F S C S S L L S	2736 603
GOTCACTACCTCCGCCGCGCGCGCGCCCCCCCTTTCCTCGCGGGAATTAGAACTATGCCGTGATCCA G H Y L H R Q K D P 3 3 F L R E I R Y I A 3 D P	1008 307	CASSICITITISSEANCETEACTISSEASCAINCECTACAGAINATISSEANCECTTCAAGAINACACACCE Q V F G K L T A S S D S L E R H C T L Q E H T A \sim	2808 907
D R R F F F N T D E A A L T D I V D A L G D R	1010 131	CHARACTERICATIONATATERICACETECTCTCTTCTTCTTCTTCTCTCTCTCTCTCTCTCT	2880 931
A P C L B C H A Z H E S S P G L E H S O 1 G P	1152 355	GITCHCCCATATGGGCCCTCCCHGTGGGTCCTGGCCCAGAATCAAAACCACTCTCAGGGTCCAGAACTA V H F Y G Y L F V G F G F E F K T T L R V Q N L	1952 955
ICCACTCATCGCCTAAAGGATGGGATCCTTTTTGGGATGGTGGGGGCCTATCACTGGGCTAA S T H R L K D G I L F G H V C A Y D W G G S V L	1274 379	G C Y V V S G L 1 L S A L L F A V A H G G W Y F	3024 979
TOGETTGAAGGAGGCCACCCCCTTTTCCCCCCACGAATGCCACTGGAAGAGGAGTTCCCCCGCACTGCAG W L B G G H R L F P F R H A L B D E F P F A L Q	1296 403	CTATEMETERIC CANATCAL CHARACTERISTIC CONTROL C	3096
AMCATCAGCTACTGGGTACTCGTTTCTTCCATCTTTTGCGGGGGGCCCCGCCTGTTTCTCTCT H H A A Y L G Y S V S S H L L R G G R R L Y L S	1368 427	CCTGTOCATOCAGAGGGGCTTCAACACACACACACAGTGATGGGCAATACTCAGTGTGAGGG F V H P E B L Q H T B R L B G S H T Q C Q V V R	3168
GOGGETECTEGASTTAGACATCGAGGAAAAGTCATCOCCTTCCAGCTTAAGAAAGATGGGGGTGGAGGGTT G A P R P R H R G X V I A P Q I R K D G A V R V	1440 451	TOTCHCCTTGCCACAGGGGATGAGGGGATGAGGGTATTGAGGCTGGTCACAATGATTTCC H L G O L A K G T E V S V G L L R L V H N E F	3240 1051
CCCCAGACCTCCAGGGGGAGATGGTCATACTTTGGAGGGAG	1512 . 475	FRRAKFKS LIVVS TFELGIEEGS V	3312 1075
GATGAMCARCTGATGATTATTGTGGTTGCTGCCGCATGTTCCTGCGATCCAGACACACAC	1564 -	LOLTBASEWSES LLEVVOTEPILI	3304 1099
V T V T L V G Q Q S T L T L Q G T L Q P E P P Q	1656 323	TOCHTOTOCHTCETCATAGE ACTOTIC COCAGOODTOCTCCTCCTCCTCCTTCCTTCTTCTTCTCTCTCTCT	1123
GATGETGGGTTGGCTAGGGGGTCTTGCTGATGTGATGCANGANGCTGTTGCTGATGTGCTGTGTGTGTGTGTGTGTGTGTGTG	547	K L G F F A H K K I F E B E K B E E K L E O	3520
GEOCGETTTTGGAGATGGGCCACCACGGGGCACTGTACCTGGACCCAGAGTGGGGTCAGCCCCCCCC	1800 571	PAGANAAGGETELGGANGTECTECCTGGLETATETTECHEAGAGETTGCANAAAGGAGTTTGGGT GETERGATGGGRCAAGAAGGEGCETETGGLETATETECECAGACCAGCAGCTGACTGACTTTGACTTCTG AGGGATGCTGCTGCTAGAGATGAGGTTTTACTCAGACAAGAAGAGCTGGCACCAAAACTAGCCATGCTCC	3600 3672 3744 3816
		CACCUTEGGTTCCCTCCTCGTGATCCTGGTTCCATAGCCAACACTGGGGCTTTTGTTTG	3884

FIGURE 6

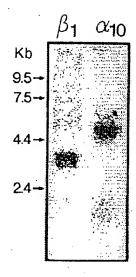
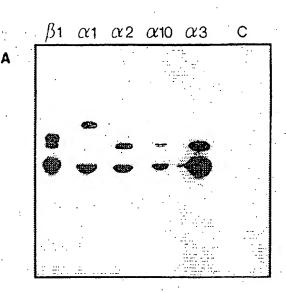


FIGURE 7

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в IP: α10 β1

Blot: β1 β1

200 -

97 -

46 -

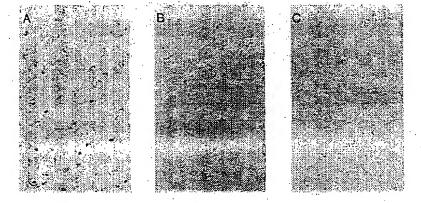
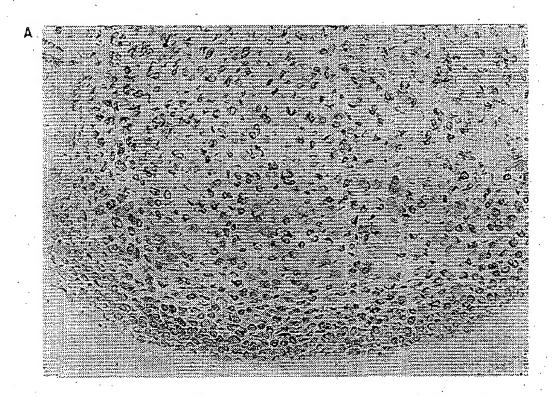


FIGURE 9



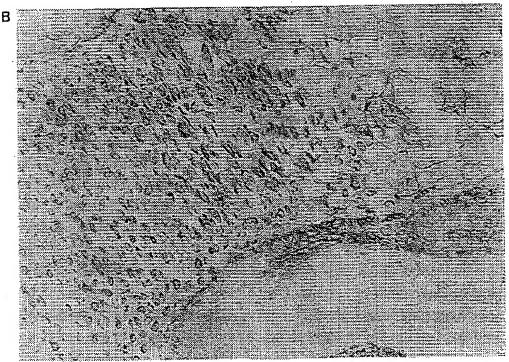
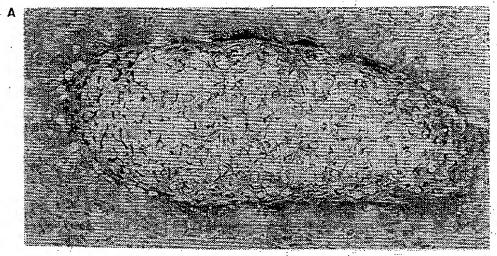
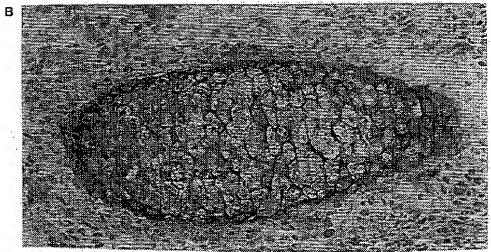
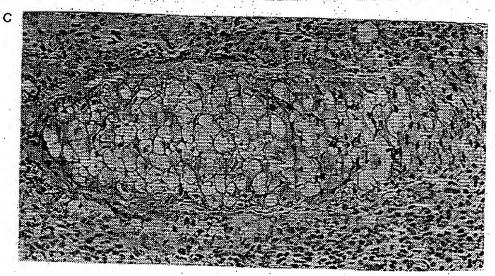


FIGURE 10

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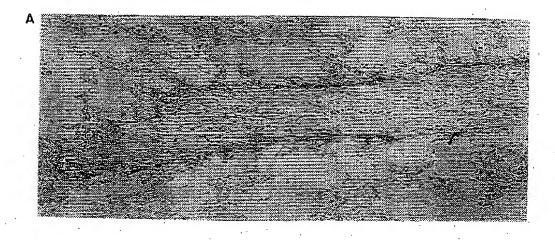


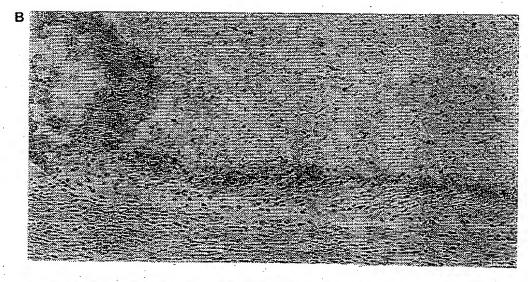
· FIGURE 11

Human RNA Master blot

Tissue	a10 expression	Tissue	$\alpha 10$ expression
Aorta	`; ; + +	Thyroid gland	-
Trachea	÷	Salivary gland	-
Lung	· ++	Spleen	•
Fetal lung	++	Fetal spleen	· * •
Kidney	++	Thymus	•
Fetal kidney	(+)	Fetal thymus	-
Heart	(÷)	Peripherial leucocyte	-
Fetal heart	++	Lymph node	•
Spinal cord	· ++.	Appendix	• •
Mammary gland	(+)	Placenta	•
Bone marrow	(.+)	Whole brain	•
Small intestine	(+)	Fetal brain	•
Skeletal muscle		Amygdala	-
Liver		Caudate nucleus	-
Fetal liver	• •	Cerebellum	-
Colon	. - '	Cerebral cortex	. - .
Bladder	•	Frontal lobe	•
Uterus	-	Hippocampus	-
Prostate	• •	Medulla oblongata	-
Stomach	-	Occipitial lobe	. •
Testis	-	Putamen	• .
Ovary	-	Substantia nigra	
Pancreas	. •	Temporal lobe	-
Piutiatary gland	•	Thalamus	•
Adrenal gland		Subthalamic nucleus	-

FIGURE 12





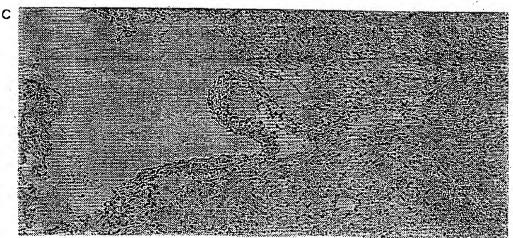
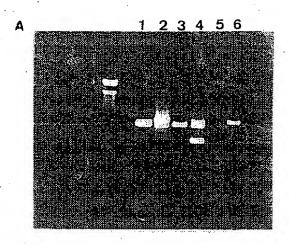


FIGURE 13



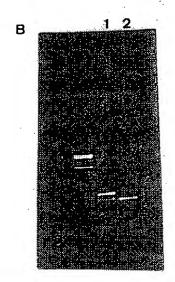
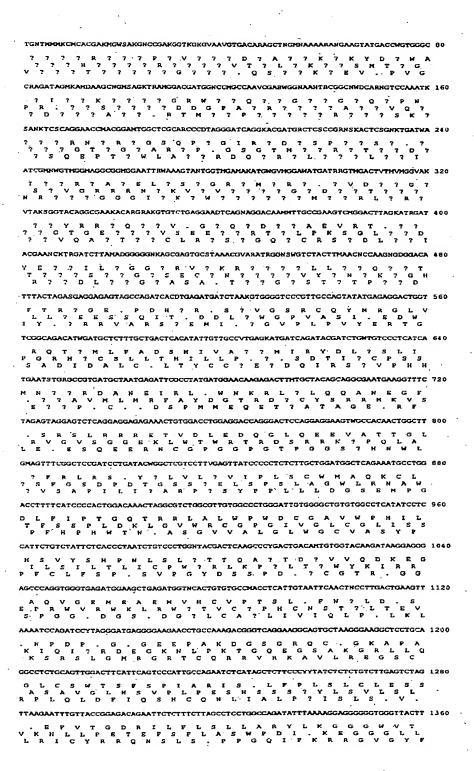
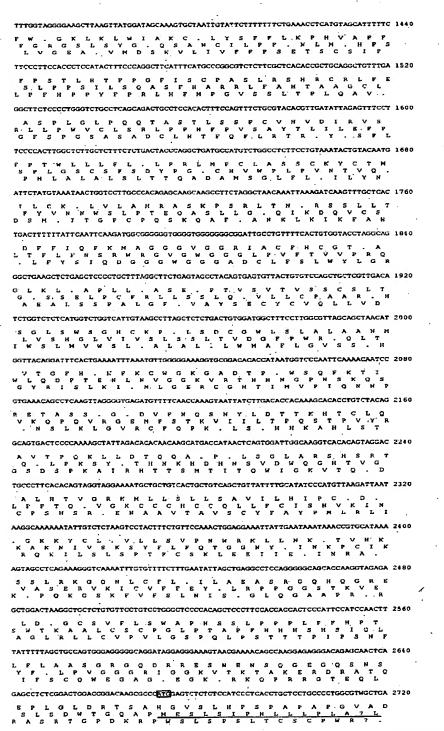
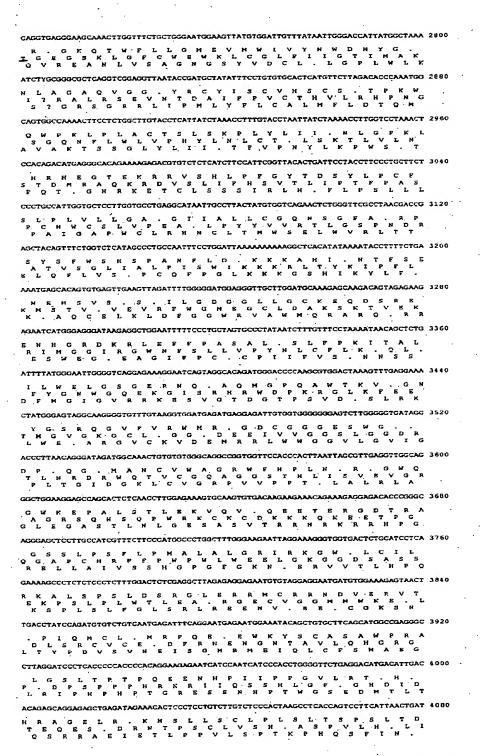
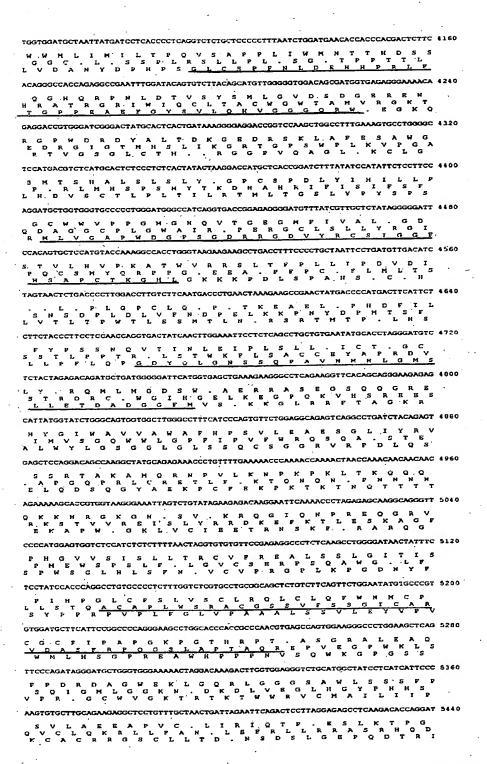


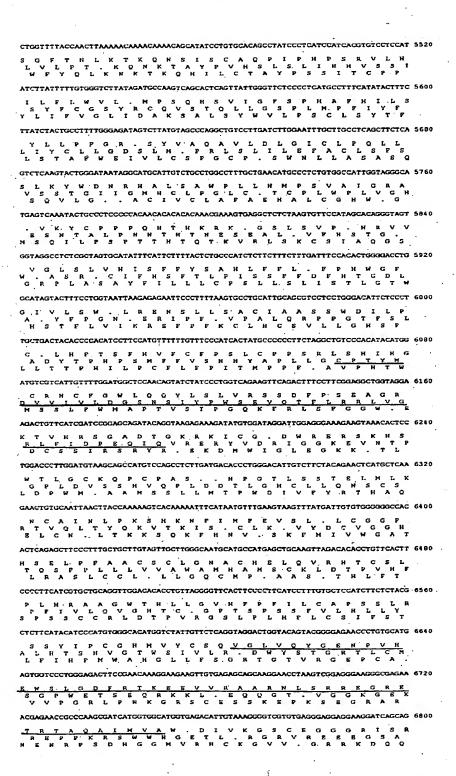
FIGURE 14











GGAGAGGGAGAGGGTCTCGAGTGTAGTGTATACATCACAAGATGCTCTGGGCGCTTATCTTTATCTGCATGCCAGAAGTT 6880	
ERERVUS VVYTS QDALGAYLYLHARS GRGRGS GV. CIHHKMLWALIFIC MPEV GEGEGLECS VYITRCS GRLS LSACQKF	
CGTGGAGGAAGGCTAGGTTGCTGTCACCATACTCTCTTACTGTATTTGCATTTTATGGTGTGTGT	
S W R K A R L L S P Y S L L L Y L H P M V S V G V S L R G G R L G C C H H T L S Y C I C I L W C L W V Y L S V B E G . V A V T I L S L T V P A F Y G V C G C I S	
CTTGTCTGTTCTGTTTCTGCACACAGAACTCCATCTTTCCTCTTCTACTCCTGCGTCAATTCTGATACCTAGCTTCTCAA 7040	
L V C S V S A H R T P S F L F Y S C V N S D T . L L N L S V L F L H T S L H L S S S T P A S I L I P S F S P C L P C F C T Q N S I F P L L L L R Q F . Y L A S Q.	
CCACTCACGCCCTAGTATTCTTTTCAAACATGACTCTAAACCTCTGGGGAGGCTACATGACCTGACTGTCTTTATTCTCC 7120	
H S R P S I L F K H D S K P L G R L H D L T V F I L T T H A L V F F S N M T L N L W G G Y M T . L S L F S P L T P : Y S F Q T . L , T S G S A T . P D C L Y S P	
AGTICCTIGATCTIGICAACCCAAGTGTITGCTGAATGAATCTATAAATAAATAATGCTTGTACATATTTACACTGATGA 7200	
Q F L D L V N P S V C . M N L . I N N A C T Y L H	
CACATTATTTTATATGTTCCGTGCCATCTAAACAGTCAAGTTGTGACTCTGTGCCAGTTTGCATGCTAGATACTGTTGGG 7280	
Q I I L Y V P C H L N S Q V V T L C Q F A C . I L L G R L F Y M F R A I . T V K L . L C A S L H A R Y C W T D Y F I C S V P S K Q S S C D S V P V C M L D T V G	
GAATGGTGTAGAAGACATCTGACCTCAGTGAACTGCTGACAGTGTTAATACACTATACGGGGCATGCCTGCATGCA	
NGVEDI.PQ.TADSVNTLYGHACHQA GMV.KTSDLSELLTVLIHYTGMPACKP EWCRRHLTSVNC.QC.YTIRACLHASL	
GTGTGTATGTGCATGCATATGCACACACACATACATATGACCATATAGCATTCTTTTATCTCTTCTTTAGCACAGAAGGGT 7440	
C V Y V H A Y A H T Y I . P Y S I L L S L F L A Q K G V C M C M H M H T H T Y D H I A F F Y L S S . H R R V C V C A C I C T H I H M T I . H S F I S L L S <u>T E G</u>	
TCAGTCAGTCCCGGGGGGGGGCGACCAGAGGCCGCTAGGCTGCTGGTAGTTGTCACTGATGGAGAGTCCCATGATGGAGAG	
S V S P G G D D Q R P L G C W . L S L H E S P M H E R O S V P G G T T R G R . A A G S C H . W R V P . W R E S O S R G G R P E A A R L L V V V T D G E S H D G E	
GAACTTCCAGCAGCGCTAAAGGCCTGTGAGGCTGGCAGAGTGACACGTTATGGGATTGCGGTGAGACTTGATCAAGTCCA 7600	
N F Q Q R . R P V R L A E . H V M G L R . D L I K S G T S S S A K G L . G W Q S D T L W D C G B T . S S P E L P A A L K A C E A G R V T R Y G I A V R L D Q V Q	
GTTGTTTTGTTTTGTGTGTATCCTCTGTGTGTGTGTGTGT	
S C F V L C C I V C V C V C V C V C V C V C V C V	
GTGTGCATGCATCAGTGCACATACCATAGTGTGTATATGCGGGTCAGAGAACCACCTCAGATGTTGGTCCTCACCTTCCA 7760	
V C M H Q C T Y H S V Y M R V R E Q P Q M L V L T P H C A C I S A H T I V C I C G S E N N L R C W S S P S C V H A S V H I P . C V Y A G Q R T T S D V G P H L P	
TCTTGTTCCAAACTGGATATCTTGTTCACTTCGGCATACAATAAGCCAGATTAGCTGACCCACAAGTCTTGGGCAGGTCT 7840	
L V P N W I S C S L R H T I S Q I S . P T S L G Q V I L F Q T G Y L V H F G I Q . A R L A D P Q V L G R S S C S K L D I L P T S A Y N K P D . L T H K S W A G L	
TCTGTCTCAGCCTCCTGTCTCTTGGTTTGAGGCATTCTGGAATTTACAGATAAGCTTGATATCGAATTCCTGCAGCCCGG 7920	
FCL SLLSLGLRHSGIYR.A.YRIPAAR SVSASCLLV.GILEFTDKLDIRFLQPG LSQPPVSWPEAFWNLQI5LISNSCSP	
GGGATCCACTAGTTCTAGAGCGGCCGCCACCAAGGGAG 7958	
GIH. P. SGRHQGS GSTSSRAAATKG GDPLVLERPPPRE	

FIGURE 15f

WO 99/51639 PCT/SE99/00544

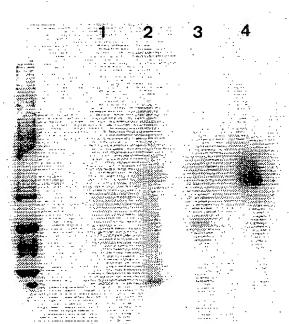


FIGURE 16

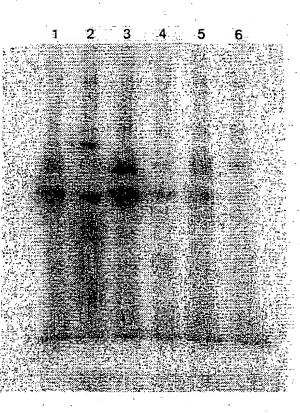


FIGURE 17

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